



# Ingénierie ostéo-articulaire et dentaire

Journée ITMO TS Rennes 2019

Pierre Weiss

INSERM U 1229

RMES Regenerative medicine and skeleton,

University of Nantes, 1 place Alexis Ricordeau, 44042 Nantes, France.

E-mail : [pierre.weiss@univ-nantes.fr](mailto:pierre.weiss@univ-nantes.fr)



## Regenerative Medicine and Skeleton research centre (RMeS)

**INSERM/UN/ONIRIS UMR 1229**

*Director Jerome Guicheux*

*Deputy-director Catherine Le Visage*

**INSERM CSS 6/CSS 3 - ITMO TECSAN/PMN**



**Nantes University Hospital Campus**

Nantes university school of dental medicine  
&  
Nantes National Veterinary School-ONIRIS



**INSERM UMR 1229-RMeS**

1700 m<sup>2</sup>-110 people

60 holding a PhD

27 clinicians (DVM, MD, DDS, PharmD)

± 20 PhD students, ± 5 post-docs

20 technical staffs

4 technological Platforms

Director J. Guicheux / Deputy-Director C. Le Visage



Scientific communication committee

Steering committee

Lab council

Emerging project initiative

## Shared services

Administrative, HR, Communication & Logistics division  
(V. Pecquetet)

Valérie Pecquetet, Tec INSERM  
Aurora Vanpoucke, Tec INSERM  
Gilles Souhiard, Ass-Tec Univ

Budget & Financial division  
(F. Seynaeve)

Fabienne Seynaeve, Ass-Eng Univ  
Carole Gauthier, Tec Univ  
Sophie Salle, Tec Univ

C. Vignes, Safety & prevention officer  
T. Rouillon, Radioactivity Competent officer

## STEP TEAM

Skeletal Physiopathology and Joint Regenerative Medicine

Coord. J. Guicheux / L. Beck (dep.)

40 PP  
27.5 FTE

5 Senior scientists  
10 Univ/hosp Researchers  
3 Post-docs  
4 Techn. Staff  
11 PhD students  
7 Master 2

Skeletal integrative physiology  
Laurent Beck, CR INSERM

Stem cells and axial skeleton development  
Anne Camus, CR CNRS

Functionalized hydrogels as joint microenvironments  
Catherine Le Visage, DR INSERM

Translational research in joint regeneration  
Jerome Guicheux, DR INSERM

## REGOS TEAM

Regenerative Medicine of Bone Tissues

Coord. P. Weiss / V. Geoffroy (dep.)

39 PP  
20.3 FTE

3 Senior scientists  
19 Univ/hosp Researchers  
2 Post-docs  
2 Techn. Staff  
5 PhD students  
6 Master 2

Molecular control of bone aging and regeneration  
Valérie Geoffroy, DR INSERM

Hybrid Biomaterials for bone scaffolds  
Pierre Weiss, PU-PH

Translational research in bone regeneration  
Florent Espitalier, PU-PH

Translational research in parodontal regeneration  
Philippe Lesclous, PU-PH

## Technical Platforms And Translational Unit

SC3M\* (J. Guicheux, C. Vinatier)

Joëlle Véziers, Eng CHU  
Julie Lesœur, Ass-Eng INSERM

Florent Autrusseau, Eng Univ  
Caroline Vignes, Eng INSERM  
Martial Masson, Eng INSERM  
Aurélie Schaefer, Tec INSERM

Cell Culture (S. Beck-Cormier)

Boris Halgand, Eng CHU

Molecular biology (F. Jehan)

Sophie Source, Tec Univ

BIO<sup>3</sup>\* (P. Weiss, C. Le Visage)

Thierry Rouillon, Eng Univ  
François Loll, Eng INSERM

Preclinical and clinical Transfer unit

1 Tec, 17 Univ/hosp researchers  
9 associated clinicians

\*SC3M: Electron Microscopy, Microcharacterization and Functional Morphohistology-imaging Core Facility

\*BIO<sup>3</sup>: Biomaterials, Biohydrogels, Biomechanics

# Bioregate, a centre of expertise in regenerative medicine

pour les sciences de la vie et de la santé

- About 200 researchers working on regenerative medicine developments in Pays de la Loire region



## Operational partners



## Financial partners



<http://www.bioregate.com/en/>





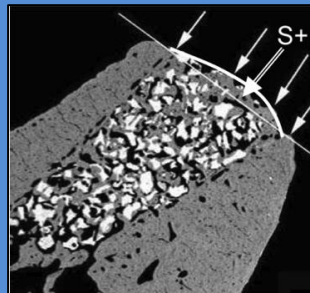
# What is regenerative medicine ?

- **Regenerative medicine** is the "process of replacing or regenerating human cells, tissues or organs to restore or establish normal function »

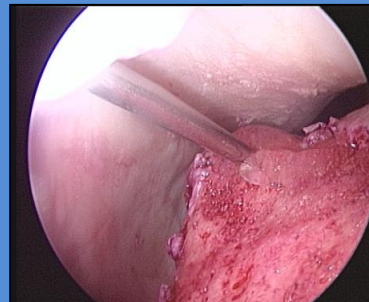
**R**eplace



**R**epair

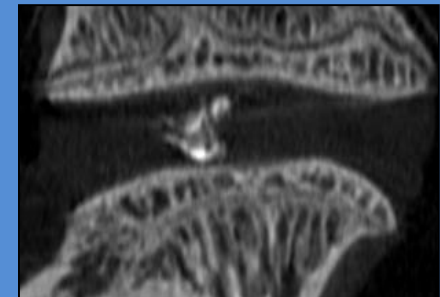


**R**egenerate



..... to **4R** medicine

**R**eprogram





# What is regenerative medicine ?

- **Biomaterials**
  - Tissue substitutes
  - Scaffold for cells attachment, proliferation and differentiation
    - Calcium phosphate
    - Polymers
    - Hydrogels
- Cellular Therapy
  - Stem cells
  - IPS
- Gene Therapy
- **Tissue engineering**

**THE NEW ERA OF REGENERATIVE MEDICINE**

*Dozens of biotech companies and university labs are developing ways to replace or regenerate failed body parts. Here are a few of the projects:*

**BONE**  
Bone-growth factors or stem cells are inserted into a porous material cut to a specific shape, creating new jaws or limbs. A product that creates shinbones is in clinical trials.

**COMPANIES:** Creative Biomolecules, Orquest, Sulzer Orthopedics Biologics, Genetics Institute, Osiris Therapeutics, Regeneron.

**SKIN**  
Organogenesis' Apligraf, a human-skin equivalent, is the first engineered body part to win FDA approval, initially for leg ulcers. Other skins are in the works for foot ulcers and burns.

**COMPANIES:** Organogenesis, Advanced Tissue Sciences, Integra LifeSciences, LifeCell, Ortec International.

**PANCREAS**  
Insulin-manufacturing cells are harvested from pigs, encapsulated in membranes, and injected into the abdomen. The method has been tested in animals and could be in human trials in two years.

**COMPANIES:** BioHybrid Technologies, Neocrin, Circe Biomedical

**HEART VALVES, ARTERIES, AND VEINS**  
A 10-year initiative to build a heart has just started. Genetically engineered proteins have been successfully used to regrow blood vessels.

**COMPANIES:** Organogenesis, Advanced Tissue Sciences, Genentech, LifeCell, Regeneration.

**SALIVA GLANDS**  
Proteins called aquaporins that allow cells to secrete water are used to recreate saliva glands damaged by disease or radiation. Glands are also being engineered to secrete healing drugs. The technique has proven successful in mice.

**COMPANIES:** None yet.

**URINARY TRACT**  
Cartilage cells are taken from the patient, packed into a tiny matrix, and injected into the weakened ureter, where they bulk up the tissue walls to prevent urinary backup and incontinence. The method is in late-phase clinical trials.

**COMPANIES:** Regeneration, Integra LifeSciences.

**BLADDER**  
Doctors at Children's Hospital in Boston have grown bladders from skin cells and implanted them in sheep. They are about to try the same process on a patient.

**COMPANIES:** Regeneration.

**CARTILAGE**  
A product is already on the market that regrows knee cartilage. A chest has been grown for a boy and a human ear on a mouse.

**COMPANIES:** Genzyme Tissue, Biomatrix, Integra LifeSciences, Advanced Tissue Sciences, ReGen Biologics, Osiris Therapeutics

**TEETH**  
Enamel matrix proteins are used to fill cavities. It works in dogs; human trials are a few years away.

**COMPANIES:** Biora, Atrix Laboratories, Creative BioMolecules.

**BREAST**  
In preclinical studies, several companies have been able to create a cosmetic nipple by inserting a ball of cartilage. Researchers are now trying to grow a whole cosmetic breast.

**COMPANIES:** Regeneration, Integra LifeSciences.

**LIVER**  
A spongy membrane is built up and then seeded with liver cells. Organs the size of a dime have been grown, but a full-size liver could take 10 years due to its complexity.

**COMPANIES:** Advanced Tissue Sciences, Human Organ Sciences, Organogenesis.

**SPINAL CORD NERVES**  
Scientists are investigating nerve-growth factors, injecting them at the site of damage to encourage regeneration or seeding them along biodegradable filaments and implanting them. Rats have been made to walk again.

**COMPANIES:** Acorda, Regeneration, CytoTherapeutics, Guilford Pharmaceuticals.

DATA: BUSINESS WEEK, DRUG & MARKET DEVELOPMENT REPORTS



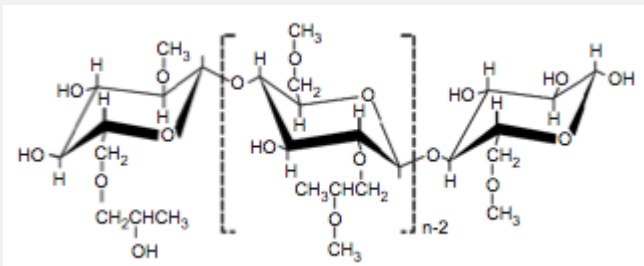
# Williams 1987

- Biomatériau :
  - Matériau non vivant utilisé dans un dispositif médical,
  - destiné à interagir avec les systèmes biologiques
  - (attention  $\neq$  matériau naturel)

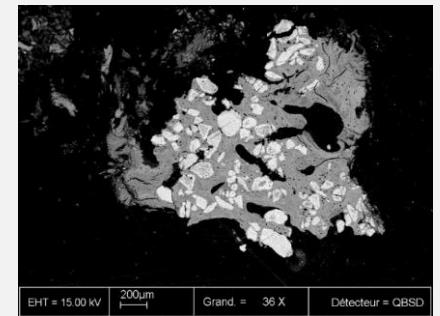
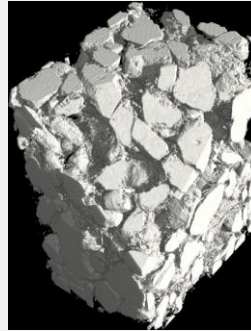




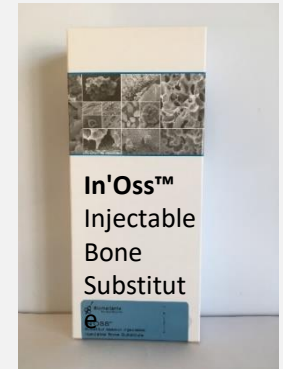
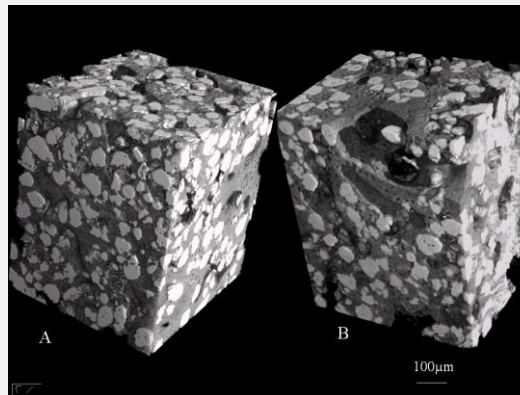
## 1992: Calcium Phosphate Suspensions in HPMC viscous water solution



2% HPMC in water + BCP granules



WO 9521634 (A1) Injectable Bone Substitute : WEISS P, DACULSI G.,  
DELECRIN J, GRIMANDI G ET PASSUTI N



Without hardening properties





pc

## Substituts osseux injectables commercialisés

### Propriétés biologiques

### Propriétés mécaniques

In'Oss™MBCP™

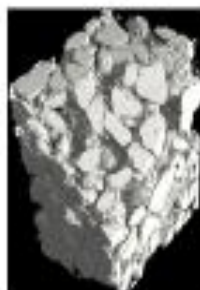
Suspension Injectable



MBCP™ Putty - In'Oss™

Hydroxyapatite + Phosphate  
Tricalcique Béta (β-TCP)  
+ viscous liquid

WD 9521634 (A1)



- + Biocompatible
- + Injectable
- + Osteoconduction
- Pas de propriété mécanique

Matériau solide

Graftys® QuickSet



Transfer



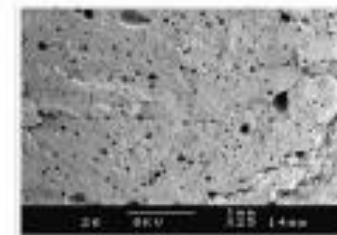
Mix 2 min



Inject

- + Biocompatible
- + Injectable
- + Propriétés mécaniques

- Porosité plus faible
- Osteoconduction



calcium phosphate salts +  
HydroxyPropylMethylCellulose  
(HPMC) + Na<sub>2</sub>HPO<sub>4</sub>

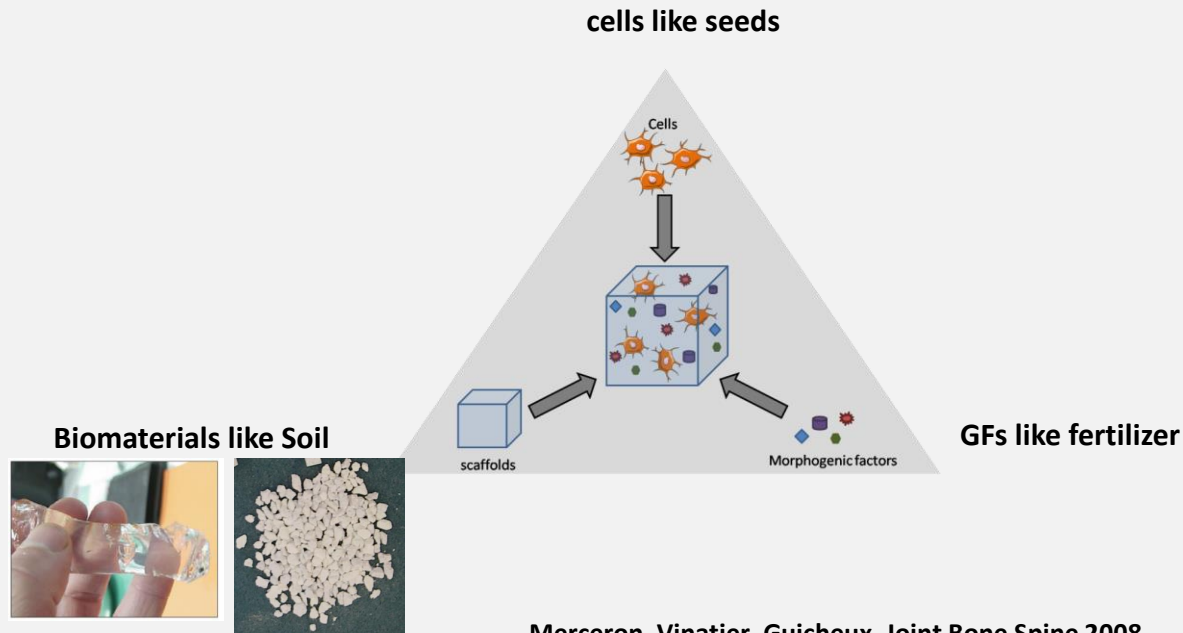


WC2008023254 A1 (2008)



# Tissue engineering

« The application of the principles and methods of engineering and life sciences toward the development of biological substitutes that restore, maintain or improve tissue function » (Woodfield, 2001).





# Why Tissue engineering?

## •2 THERAPEUTIC AXES

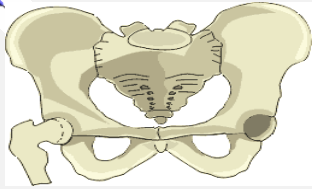
- Large bone losses.
- some affections of the cartilage



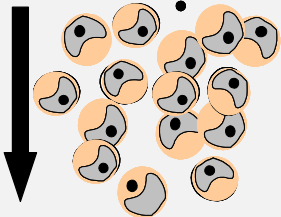




Aspiration of bone marrow or fat tissue

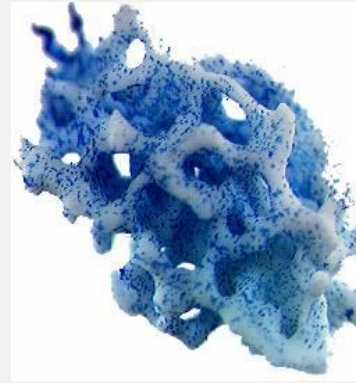


Multiplication  
 $200 \cdot 10^6$  Cel. (10-12 j)



Differentiation

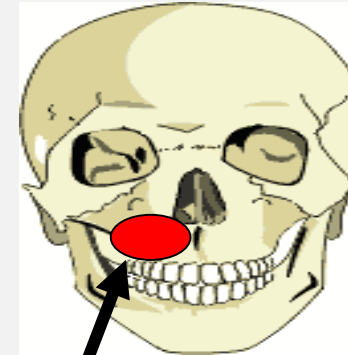
CaP



Cells + Materials



Hydrogel



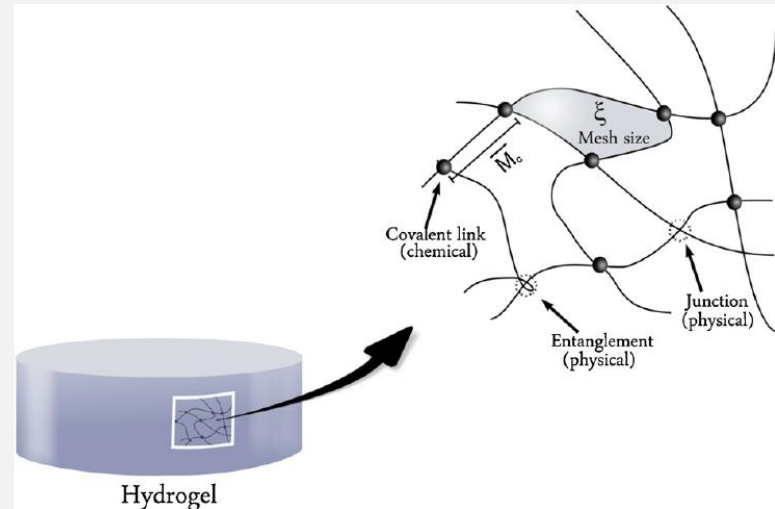
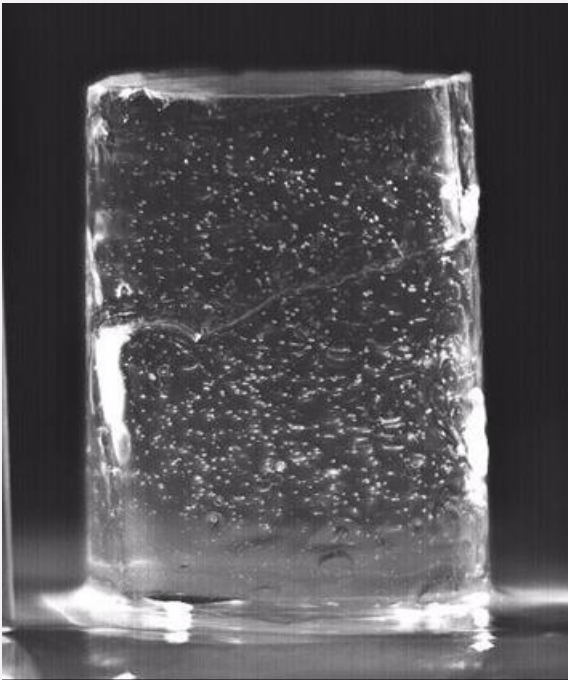
Implantation of Hybrid graft





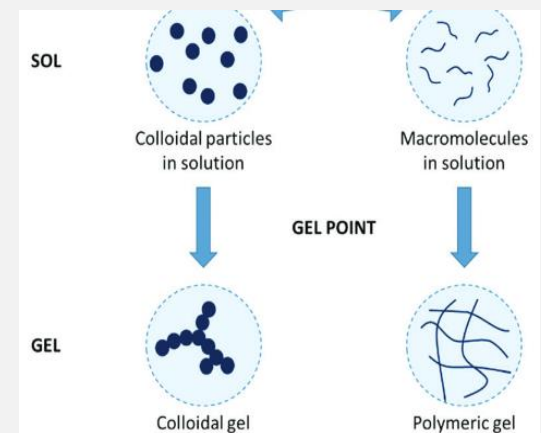


# What is an Hydrogel ?»



International Union of Pure and Applied Chemistry (IUPAC) :

"Non-fluid colloidal network or polymer network that is expanded throughout its whole volume by a fluid."



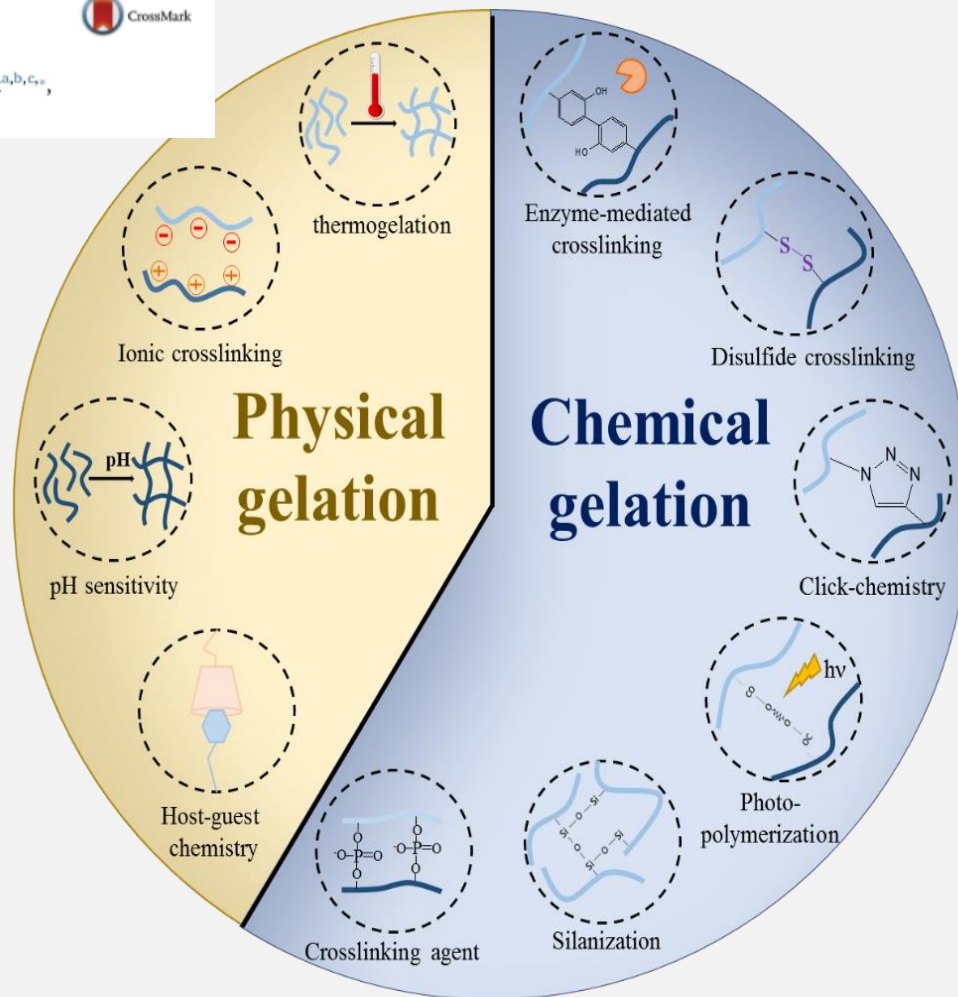


## Historical perspective

## Toward the development of biomimetic injectable and macroporous biohydrogels for regenerative medicine

Killian Flégeau<sup>a,b</sup>, Richard Pace<sup>a,b</sup>, H el ene Gautier<sup>a,b</sup>, Gildas Rethore<sup>a,b,c</sup>, Jerome Guicheux<sup>a,b,c,\*</sup>, Catherine Le Visage<sup>a,b,1</sup>, Pierre Weiss<sup>a,b,c,1</sup>

- Hydrogels in Tissue Engineering:
  - 90%+ Water
  - Hydrophilic polymer
  - Biocompatible
  - Biodegradable
  - Weak mechanical properties

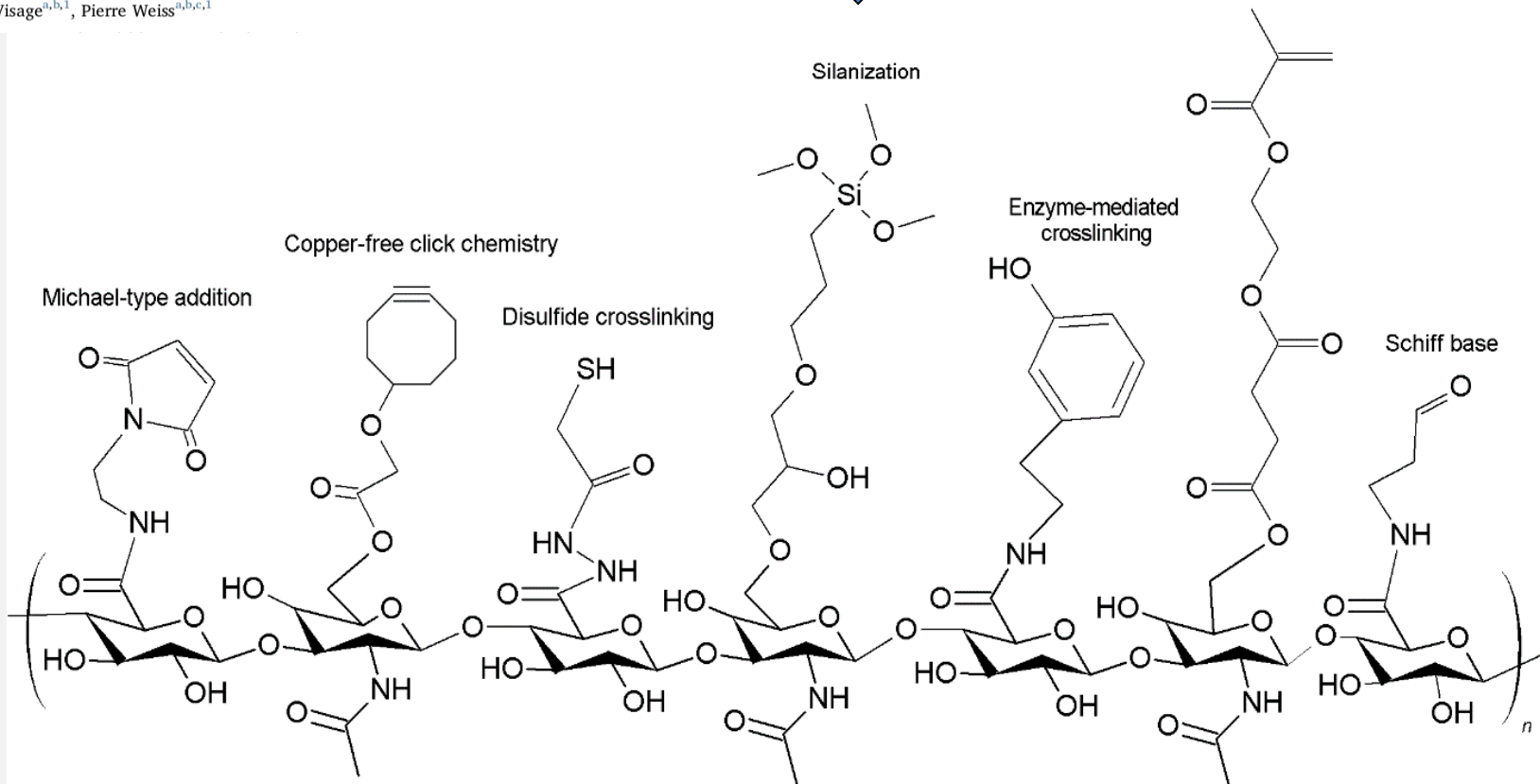




Historical perspective

Toward the development of biomimetic injectable and macroporous biohydrogels for regenerative medicine

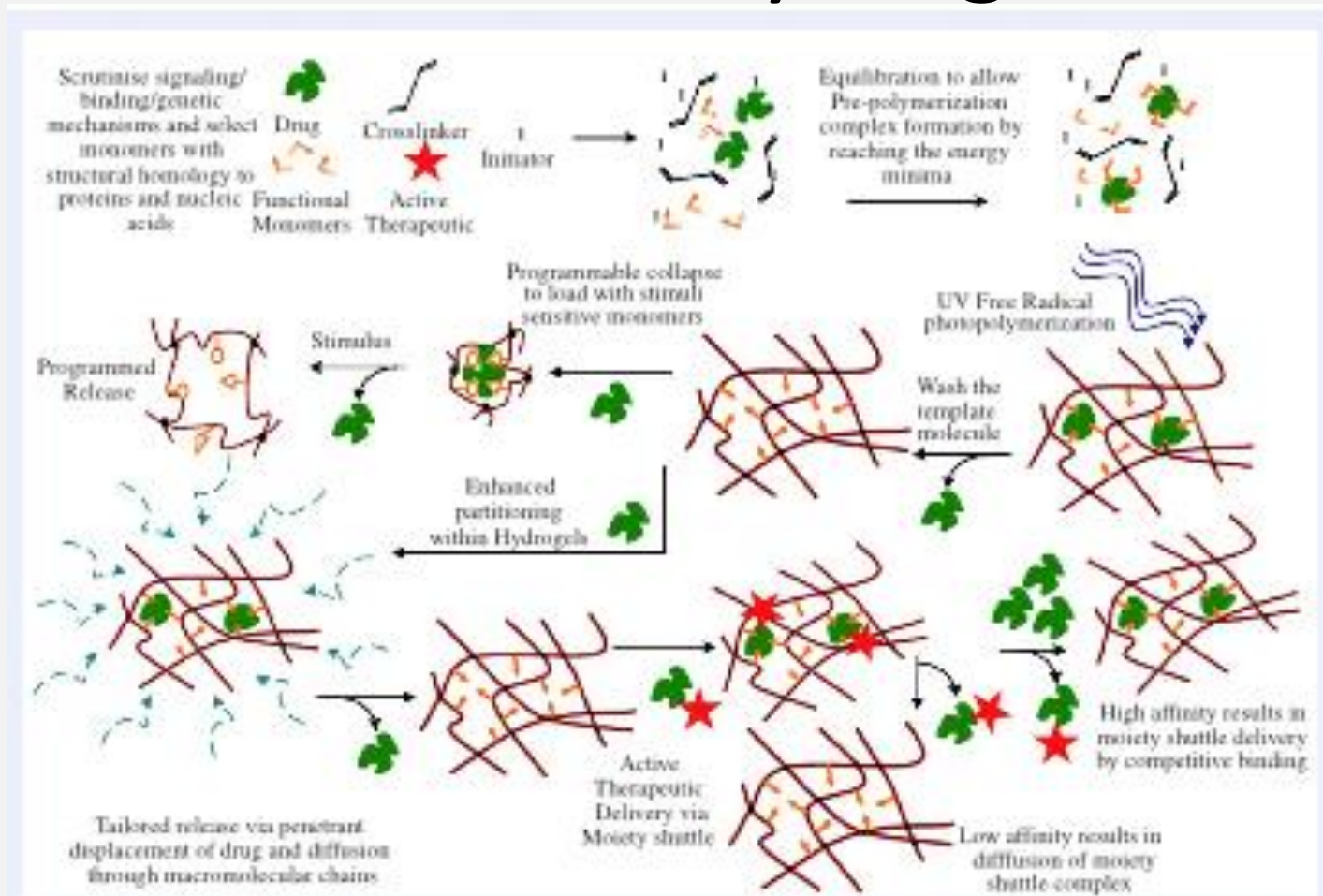
Killian Flégeau<sup>a,b</sup>, Richard Pace<sup>a,b</sup>, Hélène Gautier<sup>a,b</sup>, Gildas Rethore<sup>a,b,c</sup>, Jerome Guicheux<sup>a,b,c,e</sup>, Catherine Le Visage<sup>a,b,1</sup>, Pierre Weiss<sup>a,b,c,1</sup>



**Common chemical modifications leading to the formation of hydrogels using the example of the hyaluronic acid polysaccharide.**



# « Smart » hydrogels



Venkatesh et al. Biomimetic hydrogels for enhanced loading and extended release of ocular therapeutics. *Biomaterials* (2007) vol. 28 (4) pp. 717-24





# Cells ? and Cells interactions ?



# Hydrogels and cells

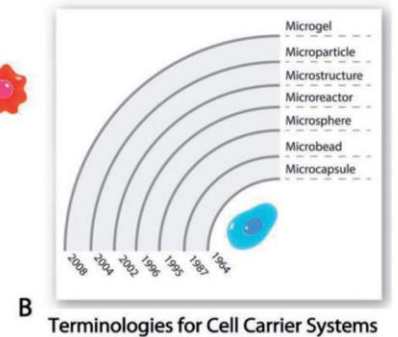
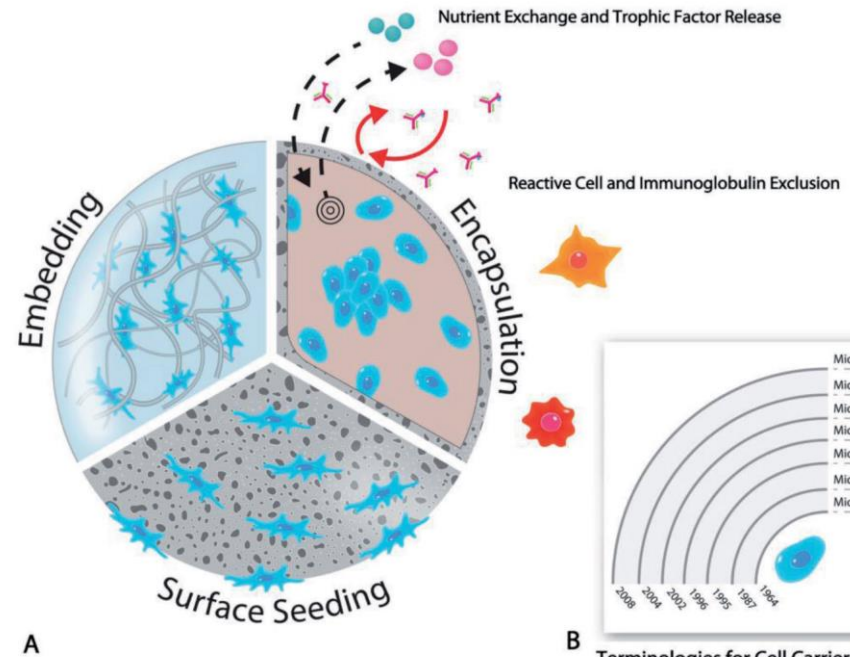
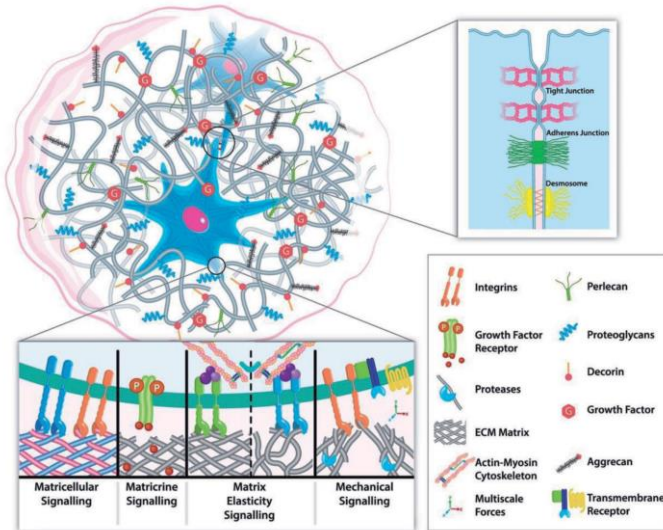
**REVIEW**

Tissue Engineering

**ADVANCED MATERIALS**  
www.advmat.de

**Toward Customized Extracellular Niche Engineering:  
Progress in Cell-Entrapment Technologies**

*Dilip Thomas, Timothy O'Brien, and Abhay Pandit\**





# Hydrogel / Cell interactions in 2D : Surface

### Alginate hydrogels as synthetic extracellular matrix materials

Jon A. Rowley<sup>a</sup>, Gerard Madlambayan<sup>a</sup>, David J. Mooney<sup>b,c,\*</sup>

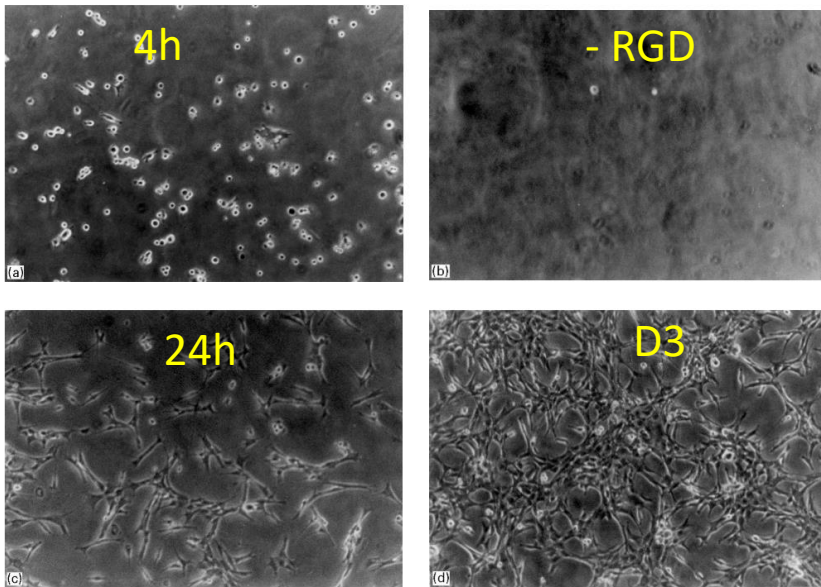


Fig. 4. Photomicrographs of myoblasts adherent to alginate hydrogel surfaces after medium changing at 4 h post-seeding on GRGDY-modified surfaces (a), and control alginate surfaces (b). The myoblasts on GRGDY-modified alginate spread extensively after 24 h of culture (c), and proliferated greatly between days one and three (d).

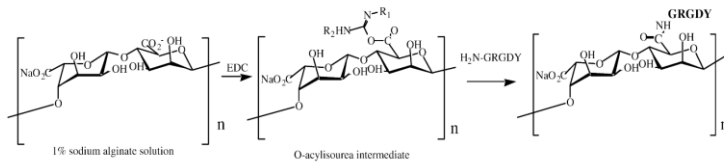
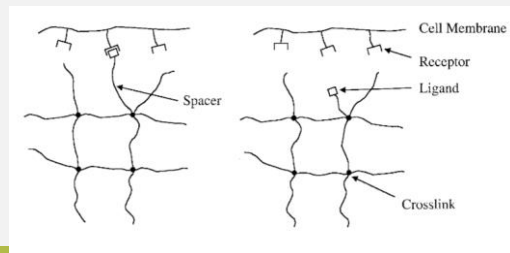
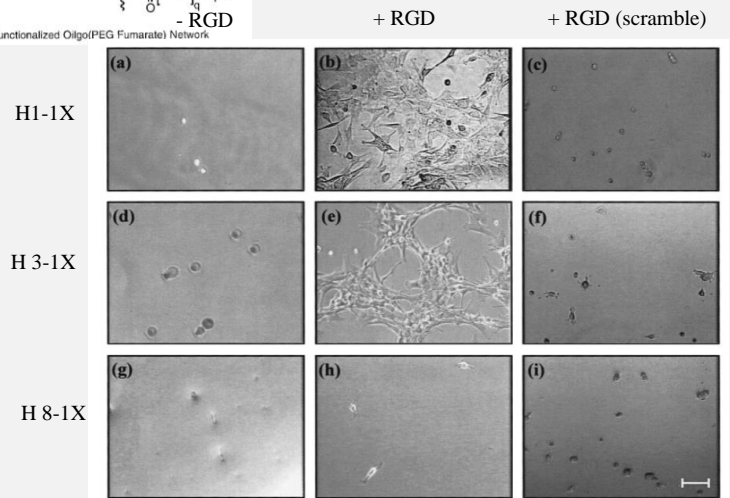
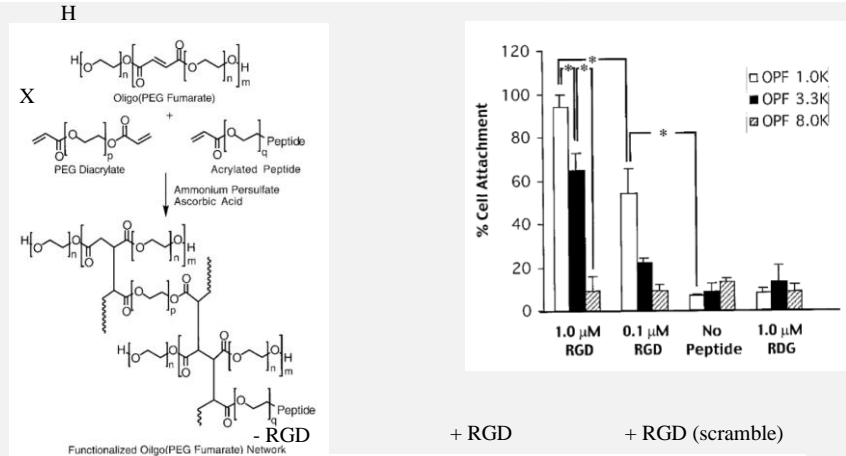


Fig. 1. Reaction scheme of peptide coupling to alginate molecules. Amide bond formation is mediated by the carbodiimide through the carboxyl group of the alginate and the N-terminal amine of the GRGDY pentapeptide.



The tripeptide Arg-Gly-Asp (RGD) consists of Arginine, Glycine, and Aspartate

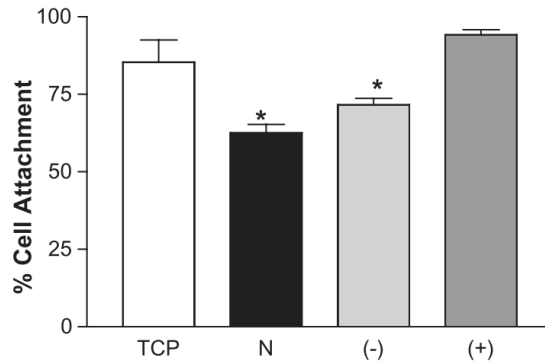




## The effect of hydrogel charge density on cell attachment

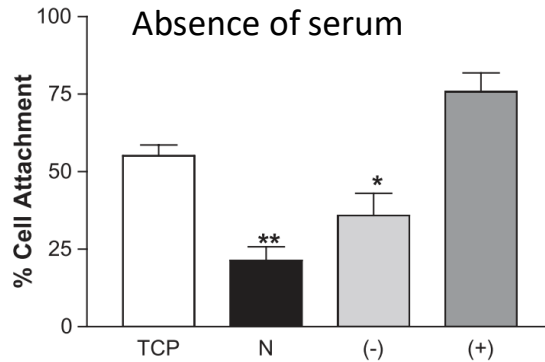
Galen B. Schneider<sup>a,\*</sup>, Anthony English<sup>b</sup>, Matthew Abraham<sup>c</sup>, Rebecca Zaharias<sup>a</sup>,  
Clark Stanford<sup>a</sup>, John Keller<sup>a</sup>

### Presence of serum

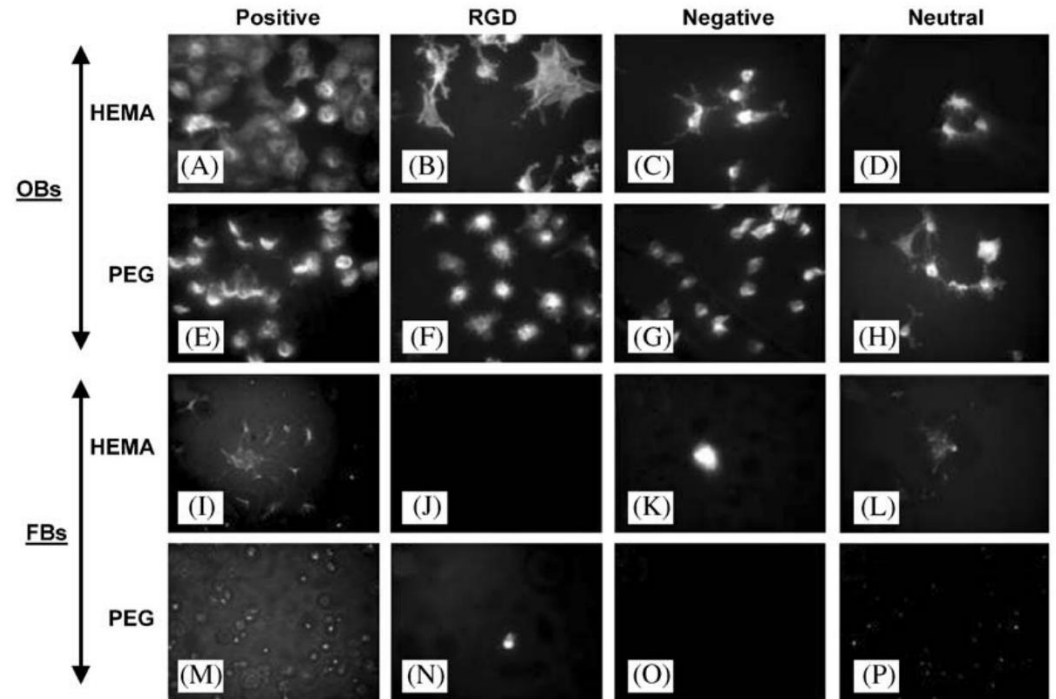


(A) Hydrogel Surface Charge Density

### Absence of serum



(B) Hydrogel Surface Charge Density



Better cell attachment and spreading was seen in osteoblasts (A–H) as compared to fibroblasts (I–P) on HEMA and PEG hydrogels with positive, negative, and neutral charge densities. Grafted RGD ligand supported cell attachment and spreading in a similar fashion as that seen on positive charge densities. Magnification, 630 $\times$ .

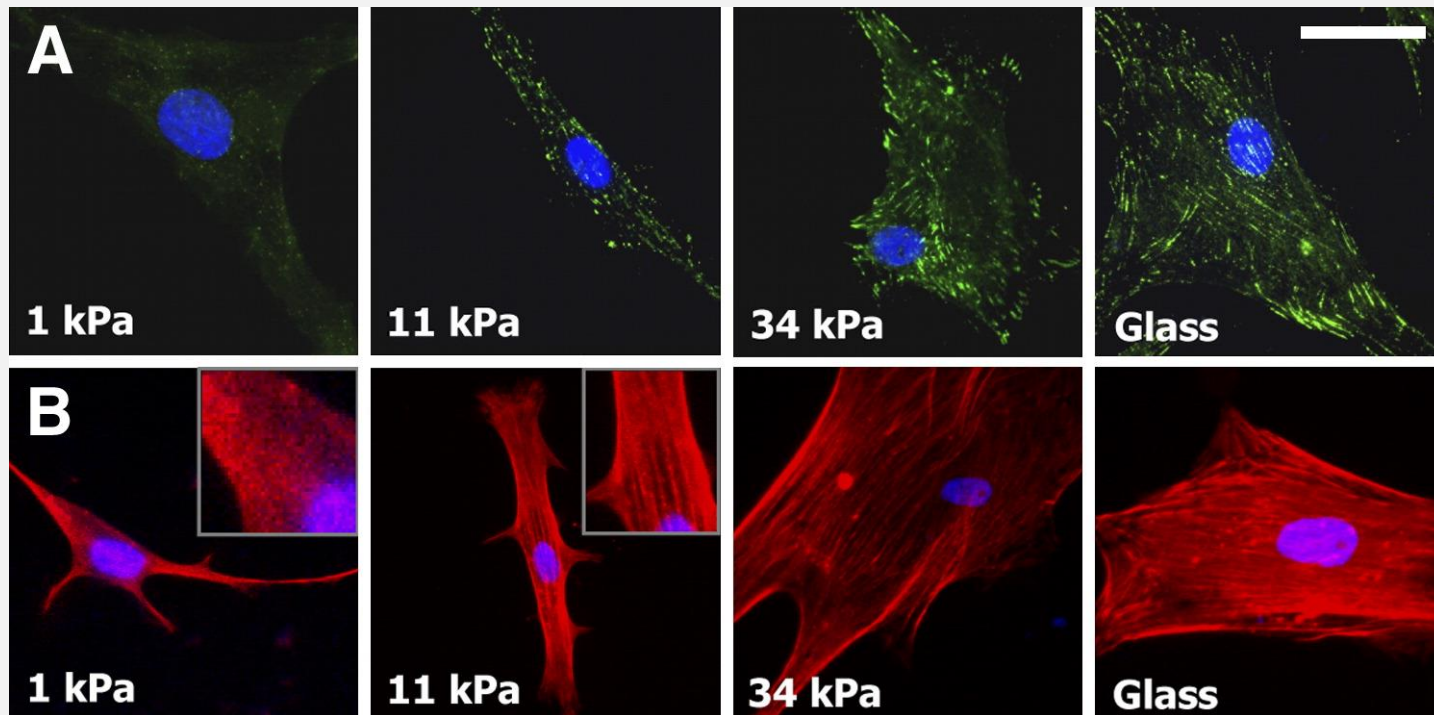


Article

## Matrix Elasticity Directs Stem Cell Lineage Specification

 Adam J. Engler<sup>1,2</sup>, Shamik Sen<sup>1,2</sup>, H. Lee Sweeney<sup>1</sup>, Dennis E. Discher<sup>1,2,3,4</sup>

# Cell adhesion



### Adhesions Grow and Cytoskeletal Organization Increases with Substrate Stiffness :

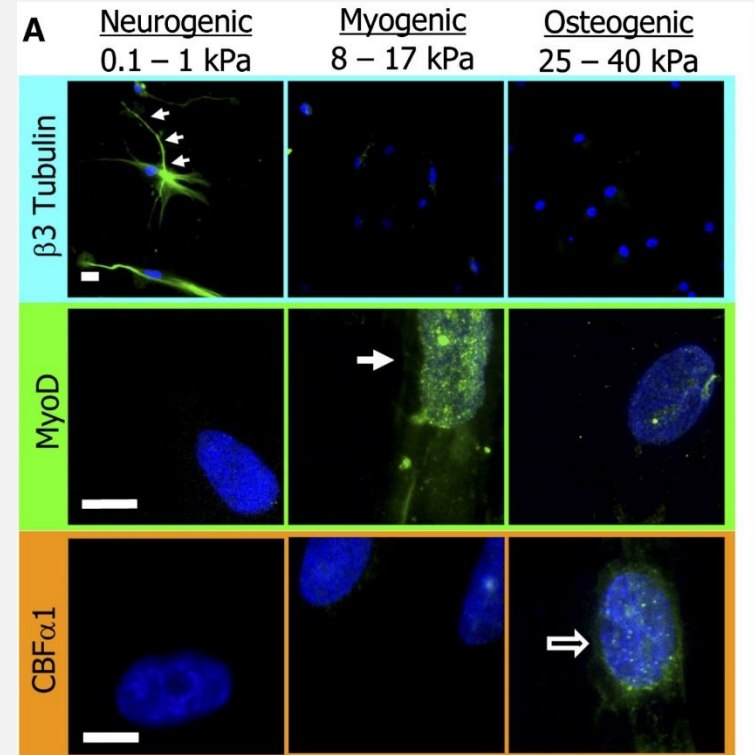
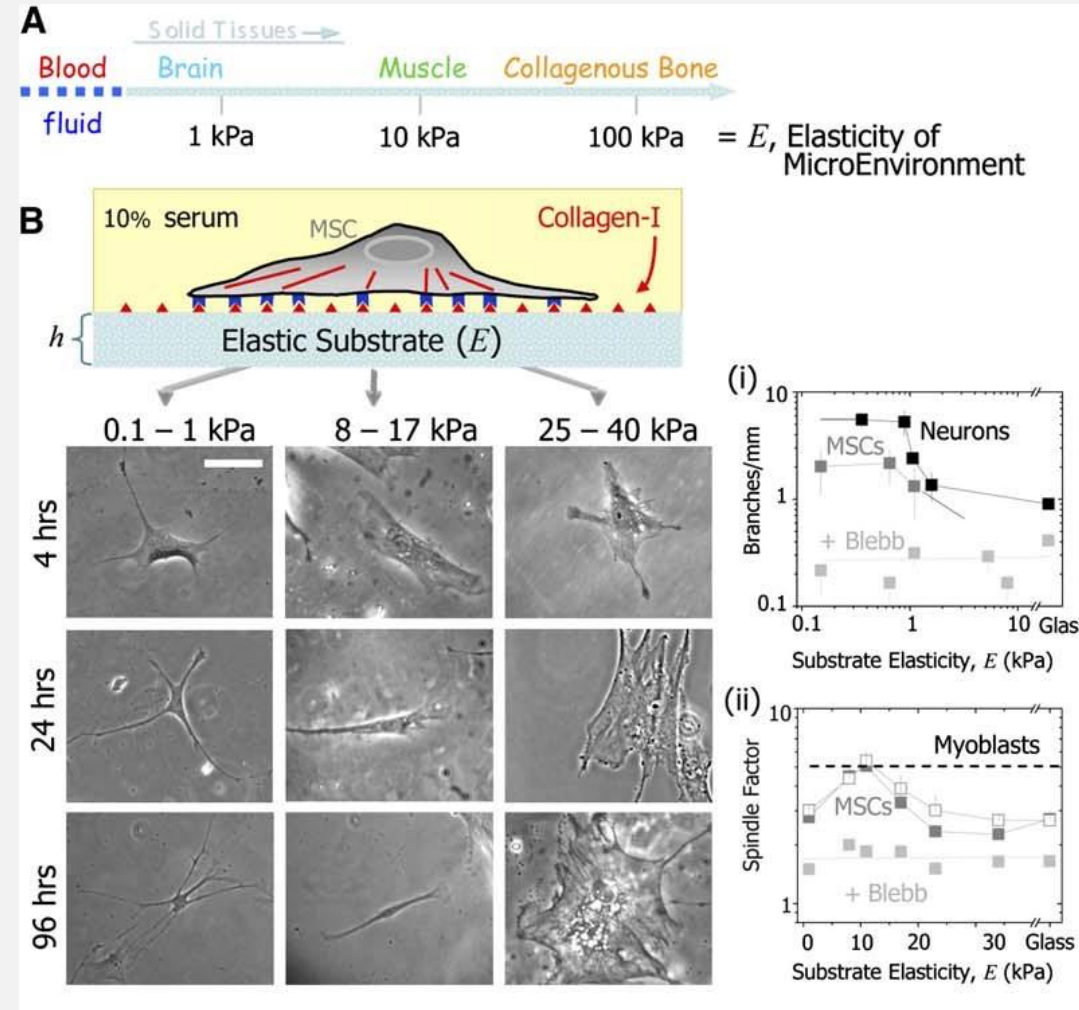
- (A) Paxillin-labeled adhesions grow from undetectable diffuse “contacts” on neurogenic, soft gels (1 kPa) to punctate adhesions on stiffer, myogenic gels (11 kPa). On the stiffest, osteogenic gels (34 kPa), the adhesions are long and thin and slightly more peripheral than they appear on glass.
- (B) F-actin organization shows a similar trend, from diffuse on soft gels to progressively organized on stiffer substrates (as stress fibers). Scale bar is 20  $\mu\text{m}$ .



Article  
Matrix Elasticity Directs Stem Cell Lineage Specification

Adam J. Engler<sup>1,2</sup>, Shamik Sen<sup>1,2</sup>, H. Lee Sweeney<sup>1</sup>, Dennis E. Discher<sup>1,2,3,4</sup> ✉

# Cell differentiation



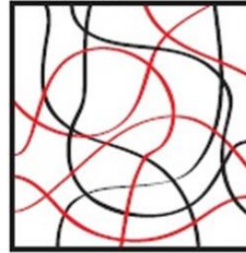




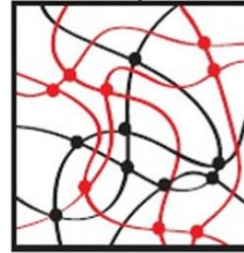
# Gel toughening?

From Peak, Wilker, et al.,  
*Colloid and Polymer Science*, 2013

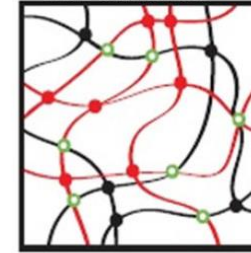
a) Interpenetrating network



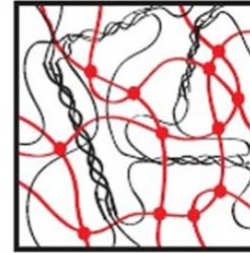
b) Double network, covalently crosslinked



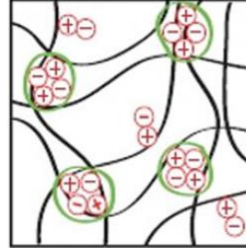
c) Double network, inter-crosslinked



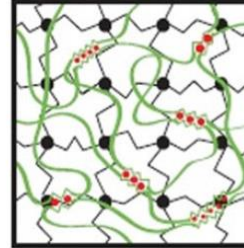
d) Synthetic biopolymer network



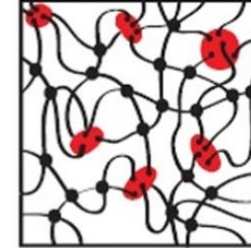
e) Ionically crosslinked network



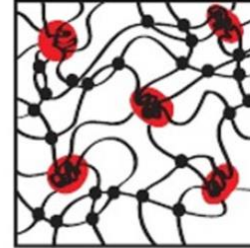
f) Ionically and covalently crosslinked network



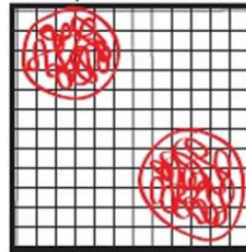
g) Double crosslinked nanocomposite network



h) Physically & covalently crosslinked network



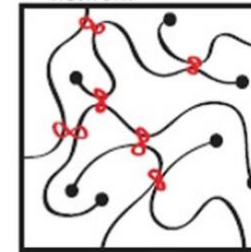
i) Temperature sensitive composite network



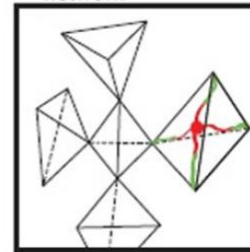
j) Composite network,  $T > T_{prep}$



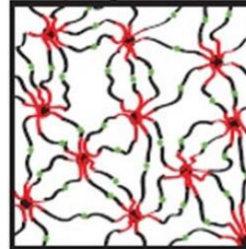
k) Slide ring hydrogel network



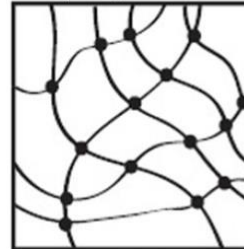
l) Tetra-PEG hydrogel network



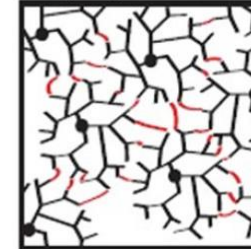
m) Micelle forming hydrogel network



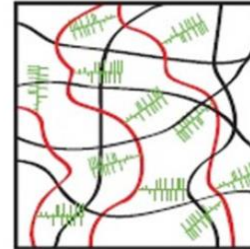
n) Single polymer hydrogel network



o) Dendritic polymer hydrogel network



p) Cartilage Schematic

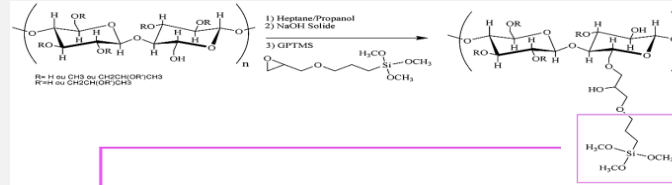






## How to reinforce hydrogels?

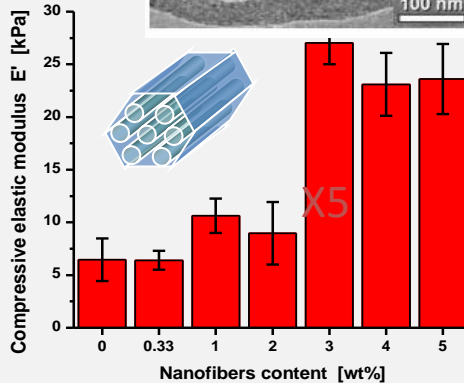
Characterization



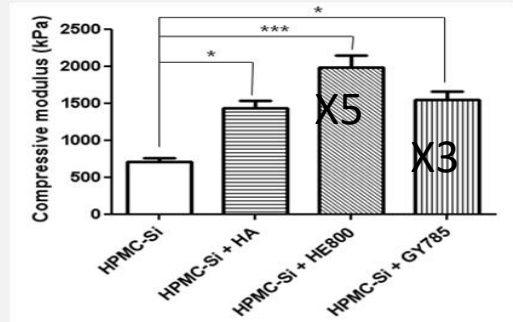
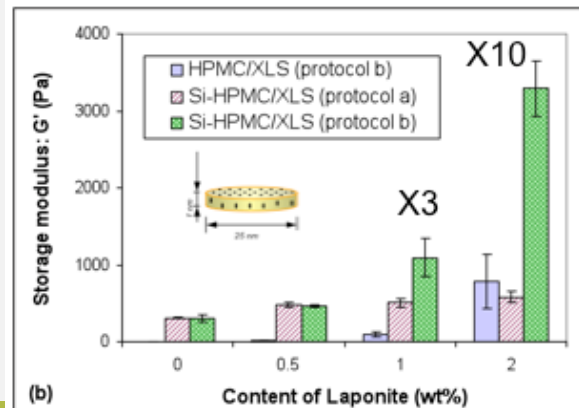
## Nano reinforcement

Marine  
macromolecules  
blended with  
SiHPMC

silica nanofibers (NFs)

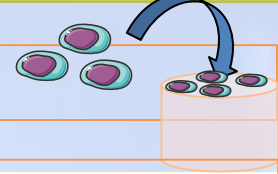


Laponite (silicate clays)



N. Buchtová,

# Cell attachment

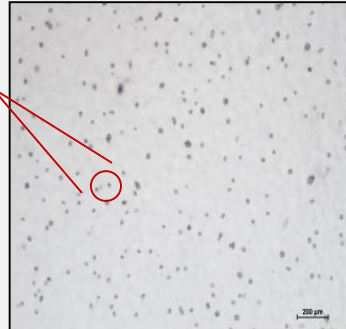


Pour les sciences de la vie et de la santé

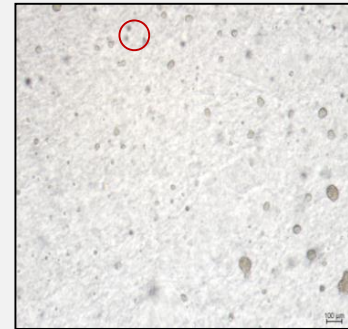
## ➤ Cell attachment at t=48h



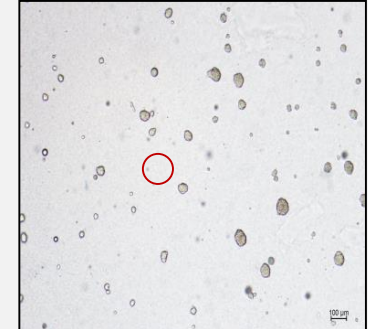
Si-HPMC



Si-HPMC+HE800



Si-HPMC+HA

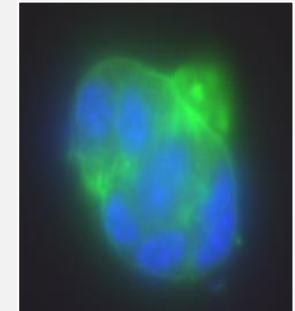
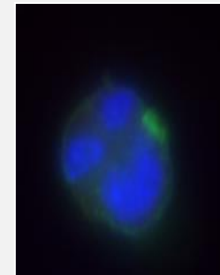
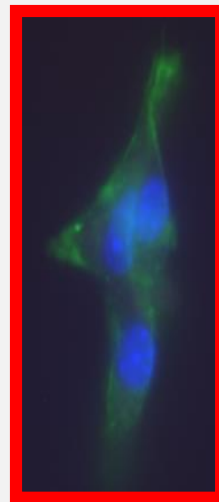


Si-HPMC+GY785

### Stainings

- Actin fibers
- Nucleus

**HE800 promotes cell spreading**



Rederstorff, E.. (2011). An in vitro study of two GAG-like marine polysaccharides incorporated into injectable hydrogels for bone and cartilage tissue engineering. *Acta Biomaterialia*.

Rederstorff, E.. (2017). Enriching a cellulose hydrogel with a biologically active marine exopolysaccharide for cell-based cartilage engineering. *Journal of Tissue Engineering and Regenerative Medicine*.

Magnification X63



# Hydrogels / Cells Interaction in 3D



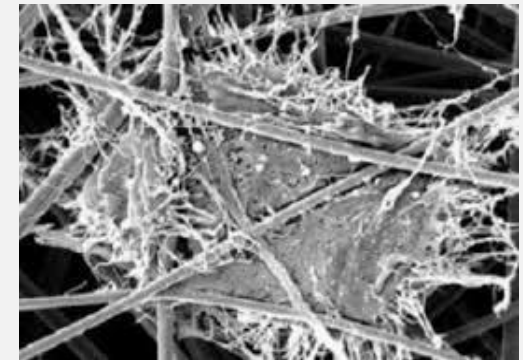
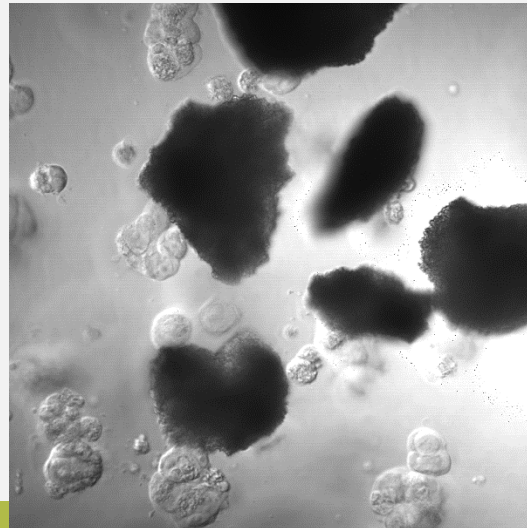
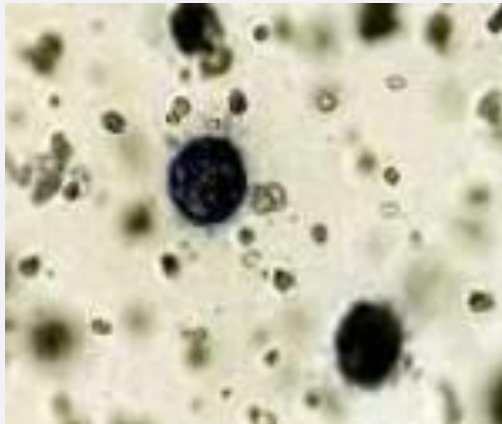
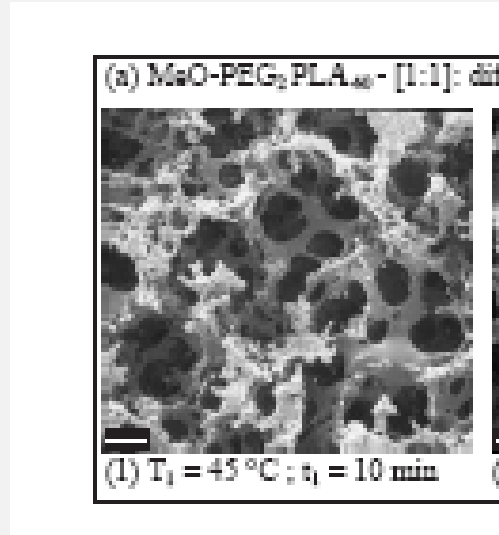
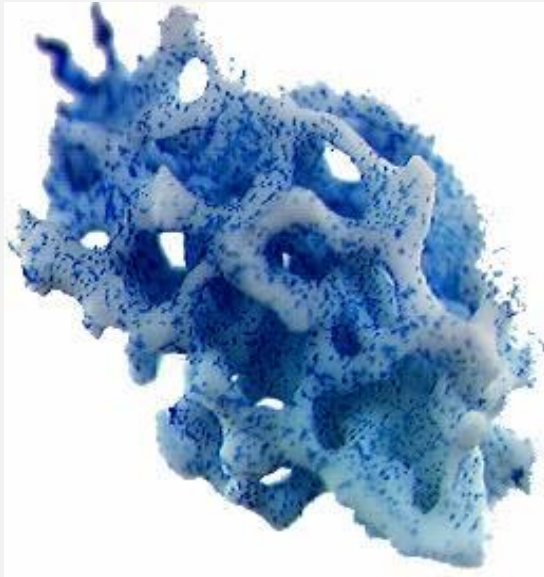
## What is 3D ?

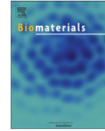






# In what type of material ?





Ac-GCRE-GPQG/IHWGQ-ERCG-NH<sub>2</sub>  
 Ac-GCRD-VPMS/MRGG-DRCG-NH<sub>2</sub>  
 Ac-GCRD-IPES/LRAG-DRCG-NH<sub>2</sub>  
 Ac-GCRD-GTAG/LIGQ-DRCG-NH<sub>2</sub>

Enhanced proteolytic degradation of molecularly engineered PEG hydrogels in response to MMP-1 and MMP-2

J. Patterson<sup>a</sup>, J.A. Hubbell<sup>a,b,\*</sup>

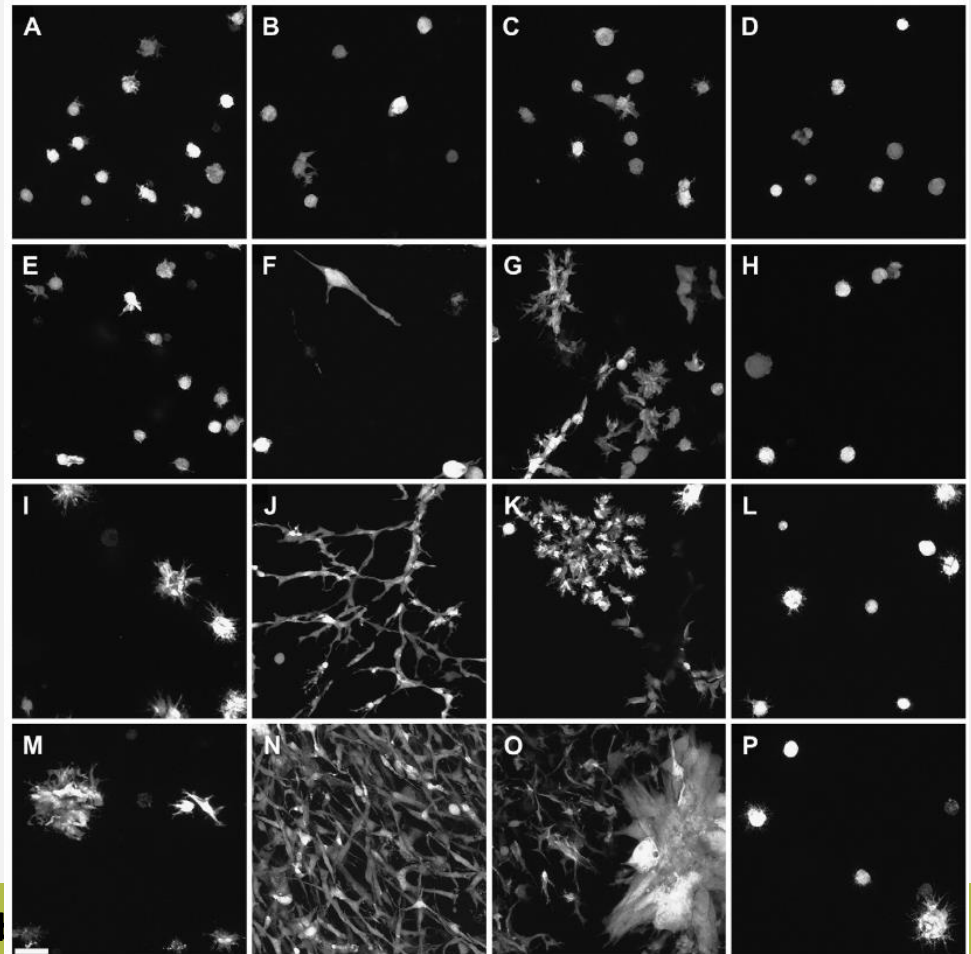
Visualization of mouse myofibroblast spreading and proliferation in 3D PEG hydrogels using confocal microscopy **with sensible peptide crosslinkers.**

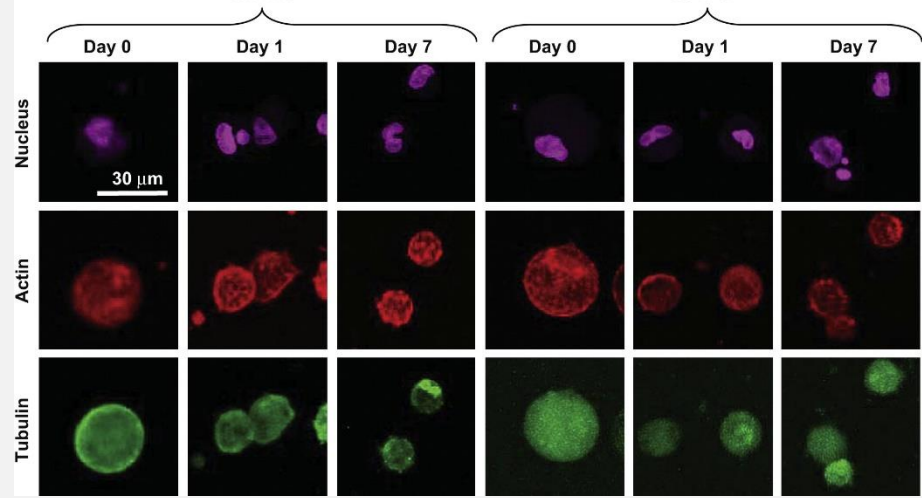
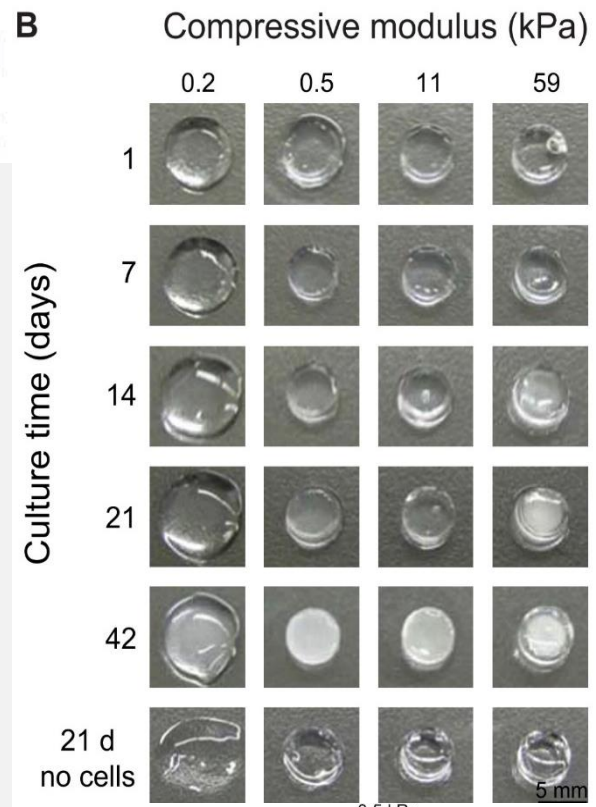
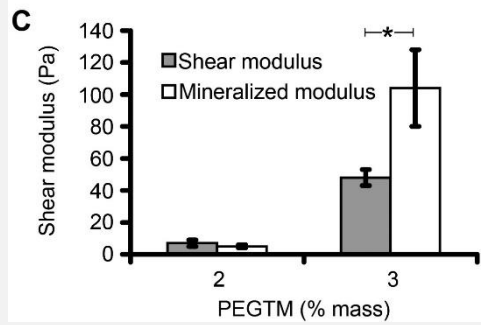
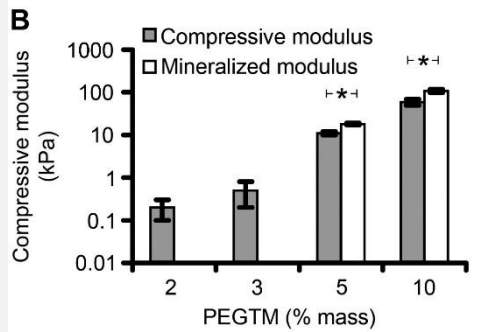
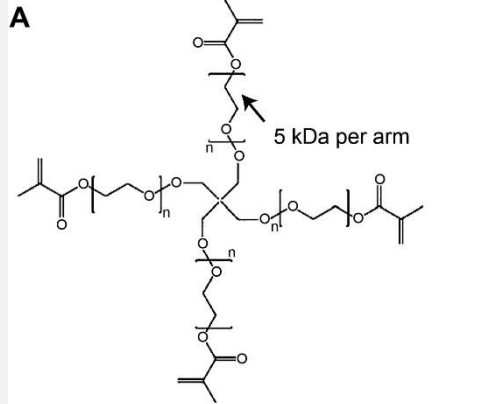
3 Days

7 days

13 Days

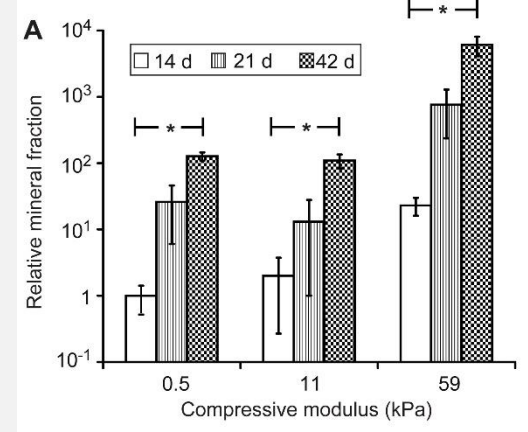
21 Days





### Modulus-driven differentiation of marrow stromal cells in 3D scaffolds that is independent of myosin-based cytoskeletal tension

Sapun H. Parekh<sup>a,1</sup>, Kaushik Chatterjee<sup>a,b,1</sup>, Sheng Lin-Gibson<sup>a</sup>, Nicole M. Moore<sup>a</sup>, Marcus T. Cicerone<sup>a</sup>, Marian F. Young<sup>b</sup>, Carl G. Simon Jr.<sup>a,2</sup>

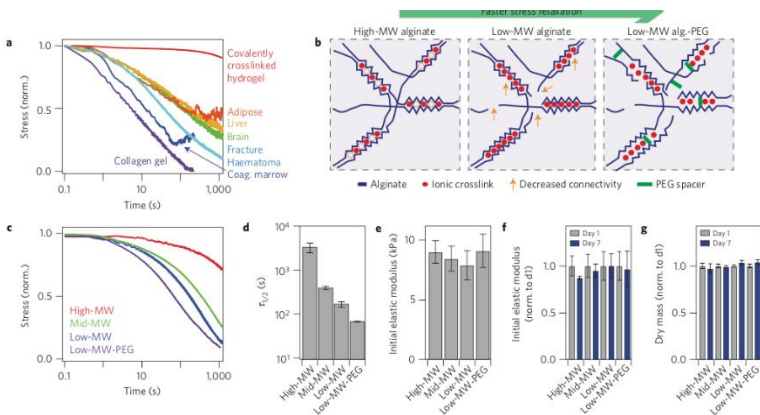




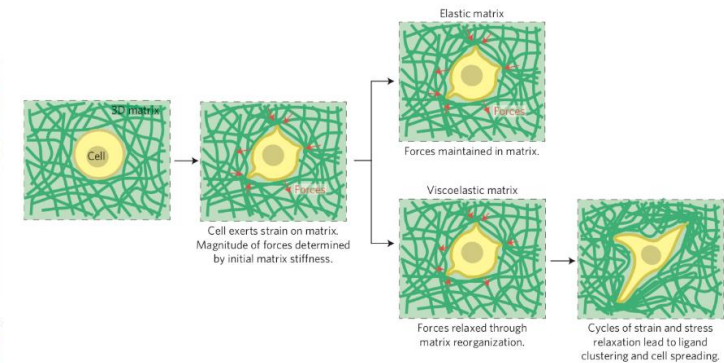
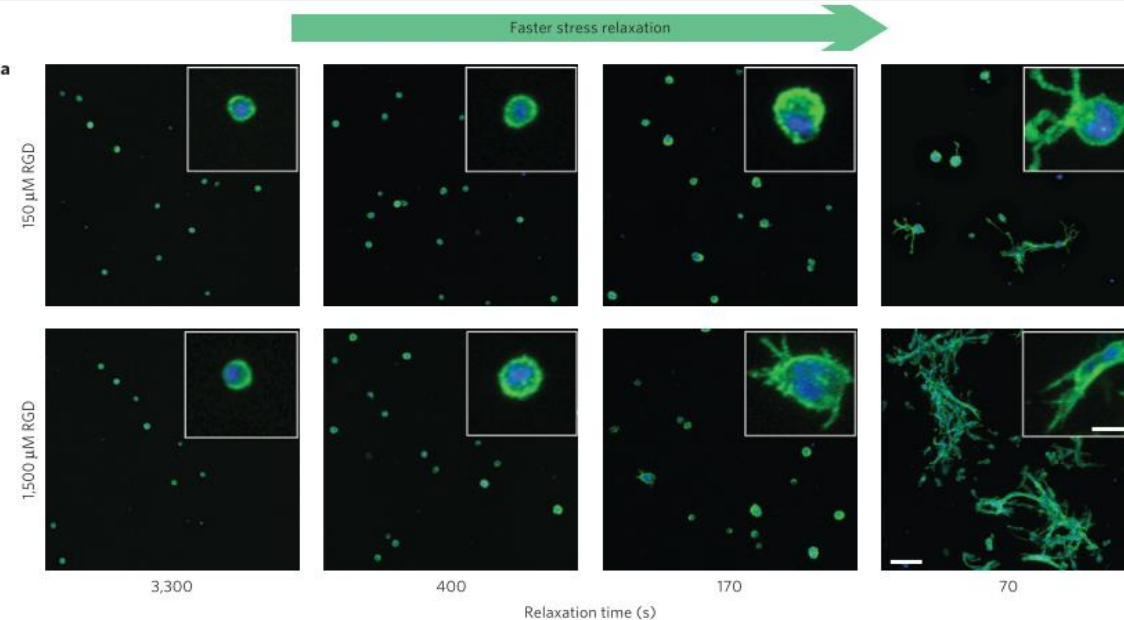
## Hydrogels with tunable stress relaxation regulate stem cell fate and activity

Ovijit Chaudhuri<sup>1,2,3†</sup>, Luo Gu<sup>1,2†</sup>, Darinka Klumpers<sup>1,2,4</sup>, Max Darnell<sup>1,2</sup>, Sidi A. Bencherif<sup>1,2</sup>, James C. Weaver<sup>2</sup>, Nathaniel Huebsch<sup>1,5</sup>, Hong-pyo Lee<sup>3</sup>, Evi Lippens<sup>2,6</sup>, Georg N. Duda<sup>6</sup> and David J. Mooney<sup>1,2\*</sup>

NATURE MATERIALS | VOL 15 | MARCH 2016 | www.nature.com/naturematerials

Cell differentiation  
In 3D

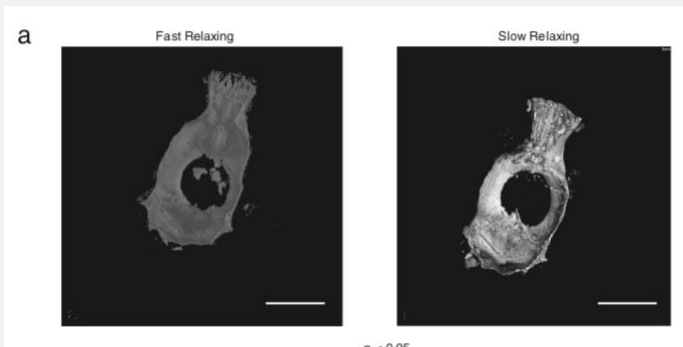
Modulating the nanoscale architecture of alginate hydrogels to modulate stress relaxation properties independent of initial elastic modulus and matrix degradation to capture the viscoelastic behaviours of living tissues



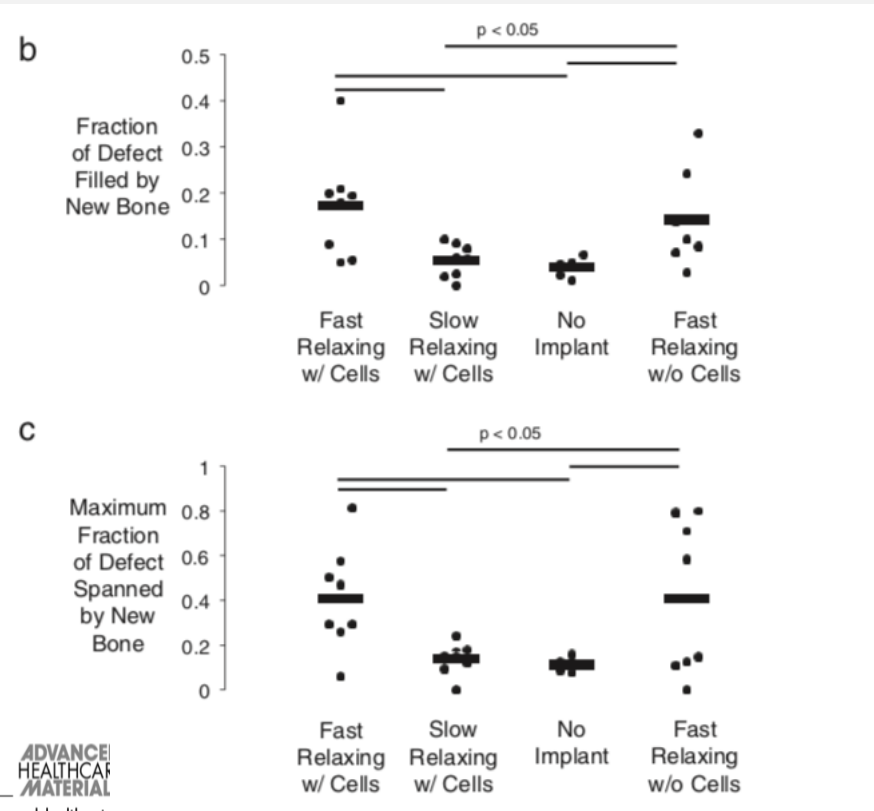




# Relaxation in vivo



**Figure 2.** Microcomputed tomography analysis of new bone formation after implantation of hydrogels in rat calvarial defect model. A) Representative CT renderings of rat calvaria three months postinjury. Scale bar, 1 cm. B) Maximum fraction of wound spanned after three months calculated by taking the maximum fraction of bone occupying any line drawn through the center of the defect. (One-way analysis of variance (ANOVA), Tukey's posthoc test,  $n = 3-4$ ) C) Fraction of the original wound area inhabited by new bone after three months. (One-way ANOVA, Tukey's posthoc test,  $n = 5-8$ ).



ADVANCED  
SCIENCE NEWS

www.advancedsciencenews.com

ADVANCE  
HEALTHCARE  
MATERIAL

www.advhealthmat.com

## Substrate Stress-Relaxation Regulates Scaffold Remodeling and Bone Formation In Vivo

Max Darnell, Simon Young, Luo Gu, Nisarg Shah, Evi Lippens, James Weaver, Georg Duda, and David Mooney\*



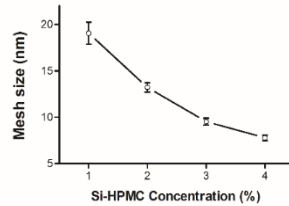
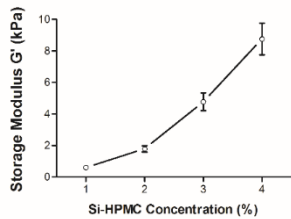
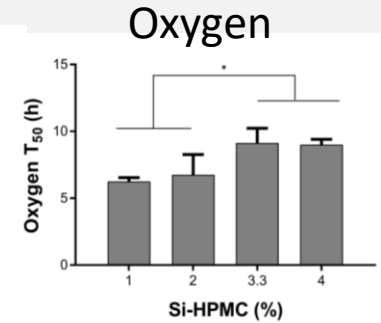
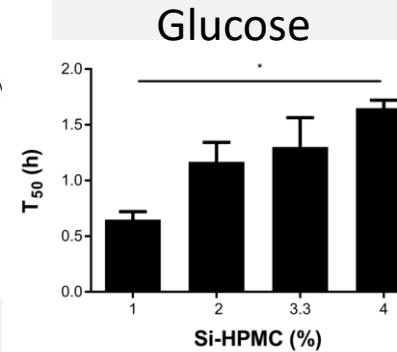
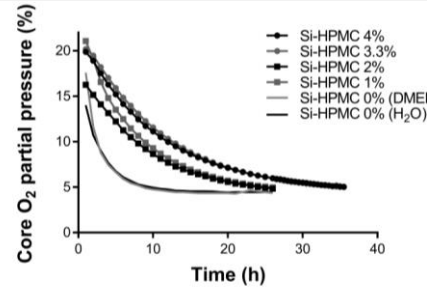
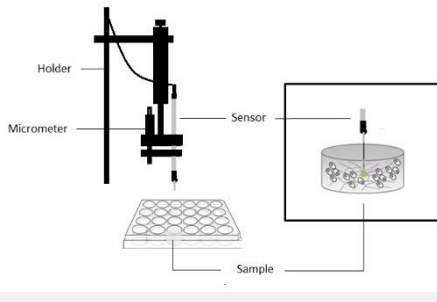
RESEARCH ARTICLE

Assessing glucose and oxygen diffusion in hydrogels for the rational design of 3D stem cell scaffolds in regenerative medicine

L. Figueiredo, R. Pace, C. D'Arros, G. Réthoré, J. Guicheux, C. Le Visage, P. Weiss

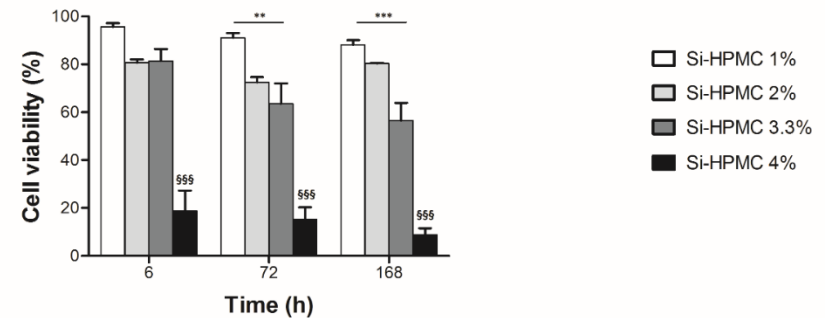
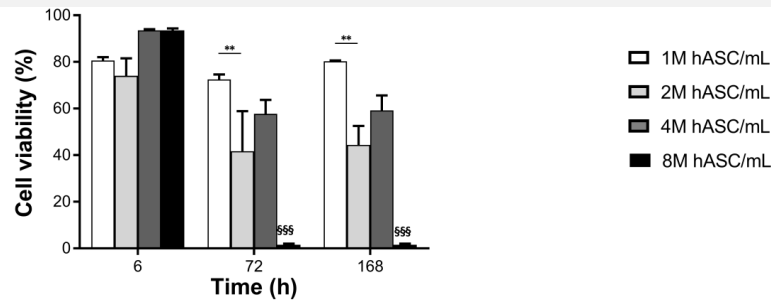
First published: 28 February 2018 | <https://doi.org/10.1002/term.2656>

# Ischemia in Hydrogels



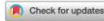
## 2% Si-HPMC

## 1 M hASC/ml



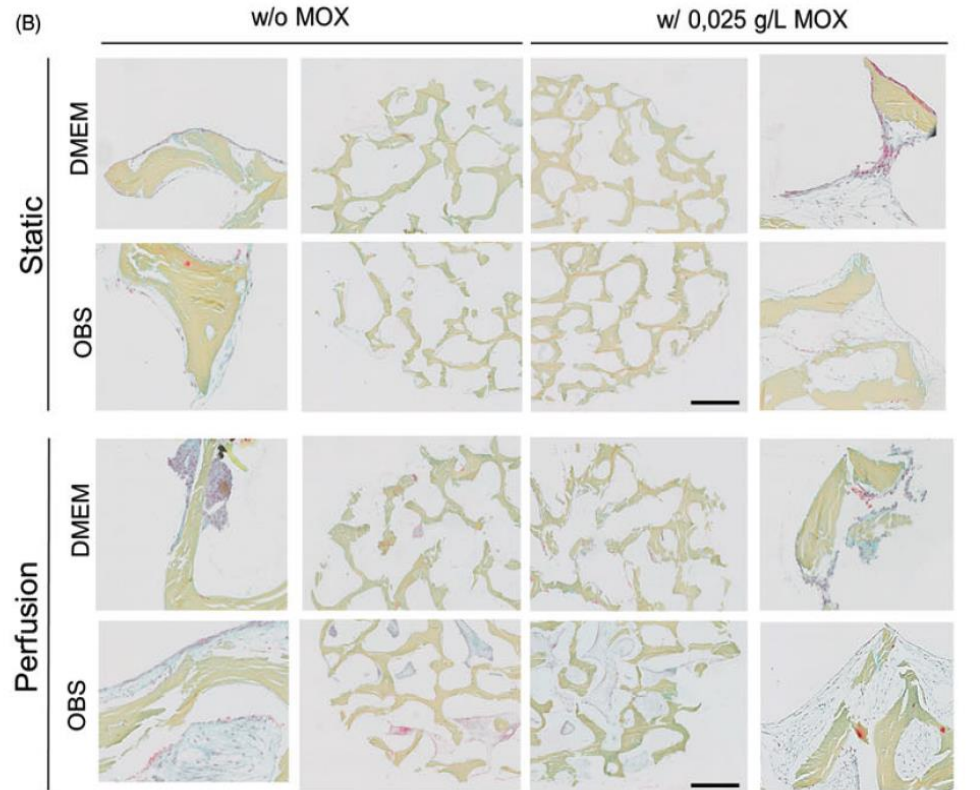
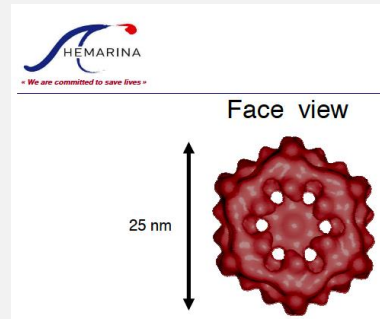
# O<sub>2</sub> and glucose in Biomaterials

ARTIFICIAL CELLS, NANOMEDICINE, AND BIOTECHNOLOGY, 2017  
<https://doi.org/10.1080/21691401.2017.1365724>



## Adhesion, proliferation and osteogenic differentiation of human MSCs cultured under perfusion with a marine oxygen carrier on an allogenic bone substitute

Fiona Le Pape<sup>a,b</sup>, Gaëlle Richard<sup>a,c</sup>, Emmanuelle Porchet<sup>a</sup>, Sophie Sourice<sup>d,e</sup>, Frédéric Dubrana<sup>f</sup>, Claude Férec<sup>a,c,f</sup>, Valérie Polard<sup>b</sup>, Richard Pace<sup>d</sup>, Pierre Weiss<sup>d</sup>, Franck Zal<sup>b</sup>, Pascal Delépine<sup>a,c,g</sup> and Elisabeth Leize<sup>a,g,\*</sup>





# Cell hydrogel interactions

- Adhesion
- Stiffness
- Electrostatic Charges
- Degradation
- Visco-elasticity
- Molecule diffusion
- Cell-cell contact
- → In 3D : you change 1 parameter all the others can change





# Hybrid Hydrogels

## REVIEW

[View Article Online](#)  
[View Journal](#) | [View Issue](#)

Check for updates

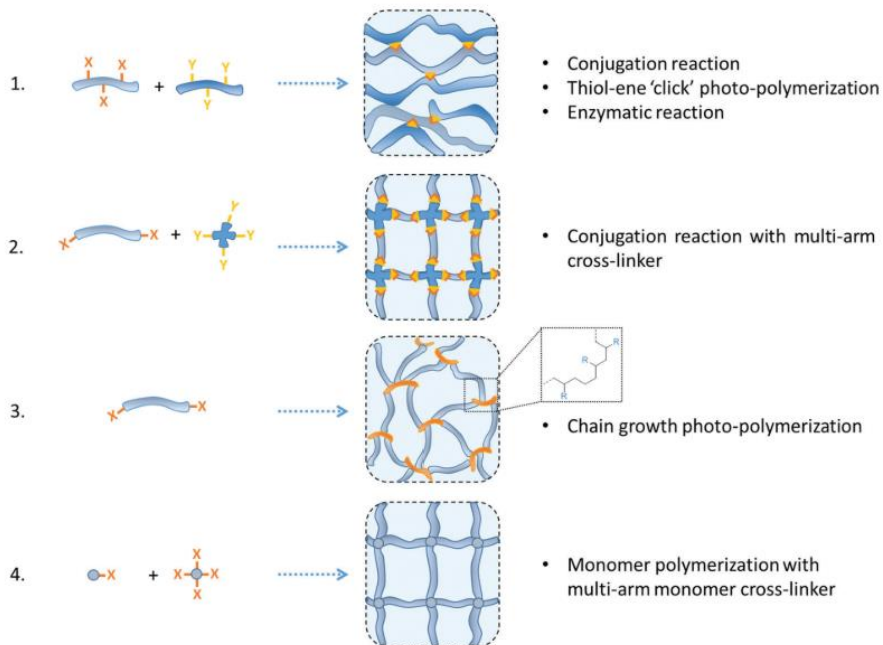
Cite this: *J. Mater. Chem. B*, 2018,  
6, 3434

## Inorganic polymerization: an attractive route to biocompatible hybrid hydrogels

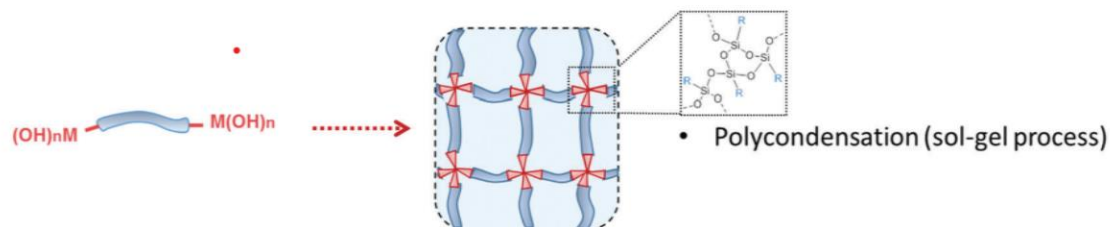
Titouan Montheil,<sup>1b</sup> Cécile Echalié,<sup>1b</sup> Jean Martinez,<sup>1a</sup> Gilles Subra<sup>1b</sup>\*<sup>a</sup> and Ahmad Mehdi<sup>1b</sup>\*<sup>b</sup>



### a. Organic hydrogel synthesis



### b. Hybrid hydrogel from inorganic polymerization



# Silated HPMC

**STEP A: Heterogeneous synthesis**

HPMC- $\text{Si}$

Alcoholic solvents

90°C

Silated HPMC powder insoluble in water

**STEP B: Solubilization and dialysis in basic water solution (pH 12,4)**

Viscous liquid phase of Si-HPMC

**STEP C: Blending liquid phases**

Si-HPMC  
pH 12,4

1 Vol

Buffer  
pH 3,6

0,5 Vol

Viscous solution  
Neutral pH  
Ambient temperature

**STEP D: Blending neutral pH viscous phase with cells and injection**

5-20 Minutes  
Polycondensation

**STEP E: Crosslink process**

Hydrogel  
Neutral pH  
 $G' > G''$

Hours

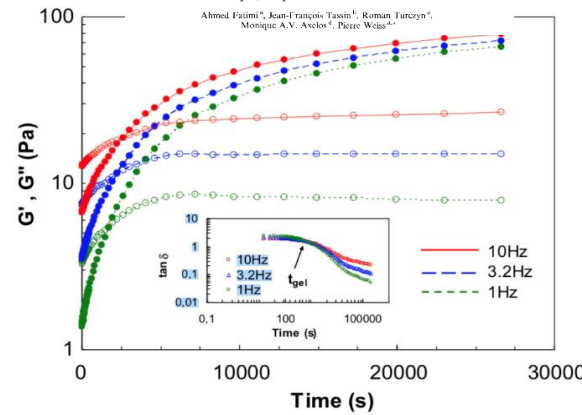
Hydrogel  
Neutral pH  
 $G'$  increases to plateau  
 $G' 300-1000 \text{ Pa}$



ScienceDirect  
ActaBIOMATERIALIA

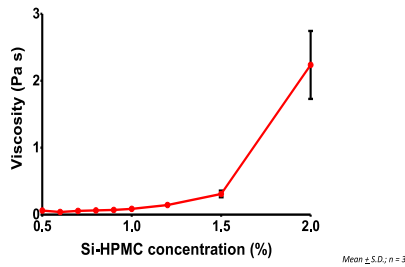
### Gelation studies of a cellulose-based biohydrogel: The influence of pH, temperature and sterilization

Ahmed Faiimi<sup>a</sup>, Jean-François Tassin<sup>b</sup>, Roman Turczyn<sup>c</sup>,  
Monique A.V. Avelos<sup>a</sup>, Pierre Weiss<sup>a,c</sup>

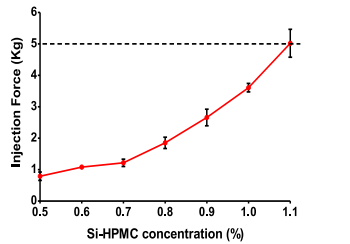


Time evolution of  $G'$  (closed symbols) and  $G''$  (open symbols) at 25°C for Si-HPMC (3%) hydrogel at pH 7.4. Three frequencies were applied (1, 3.2 and 10 Hz) at a fixed total shear stress (1 Pa). The tandisshown in the inset vs. the frequency for Si-HPMC hydrogel where gel time ( $t_{gel}$ ) is indicated.

### Rheology: Viscosity



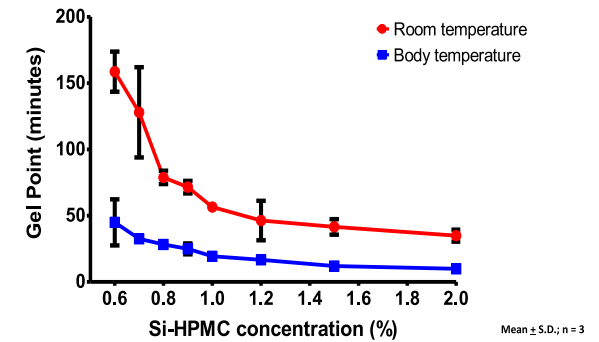
### Rheology: Injectability (Direct)



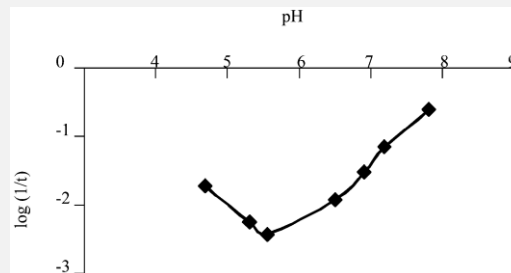
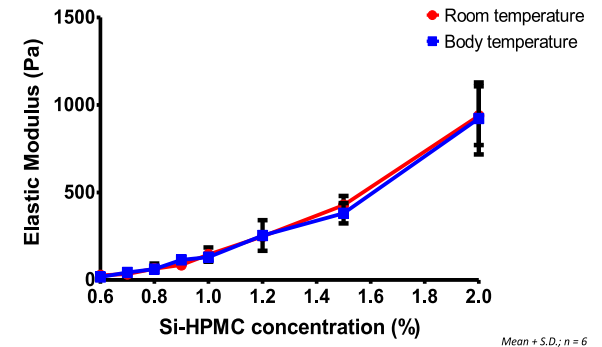
G.Pattapa, ANR Anthos



### Rheology: Gel-Point



### Rheology: G'



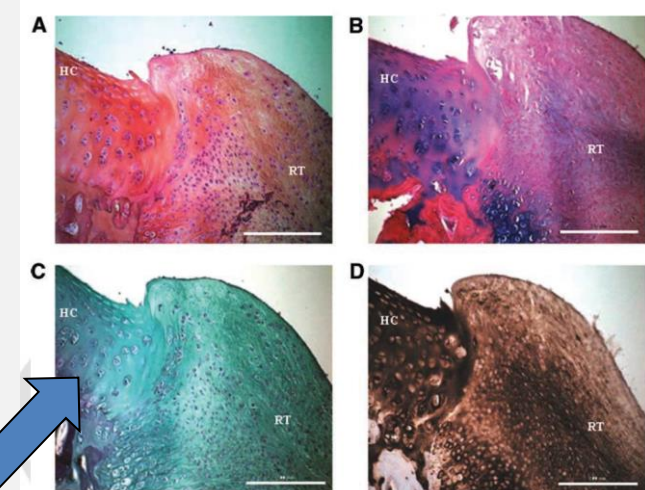
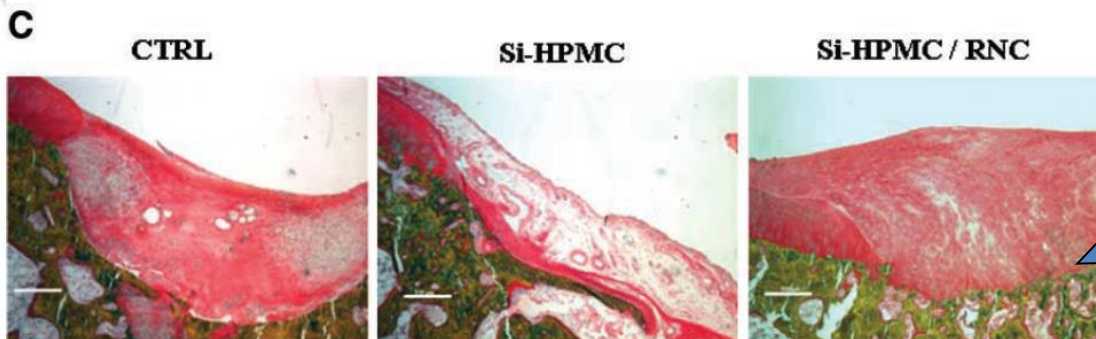
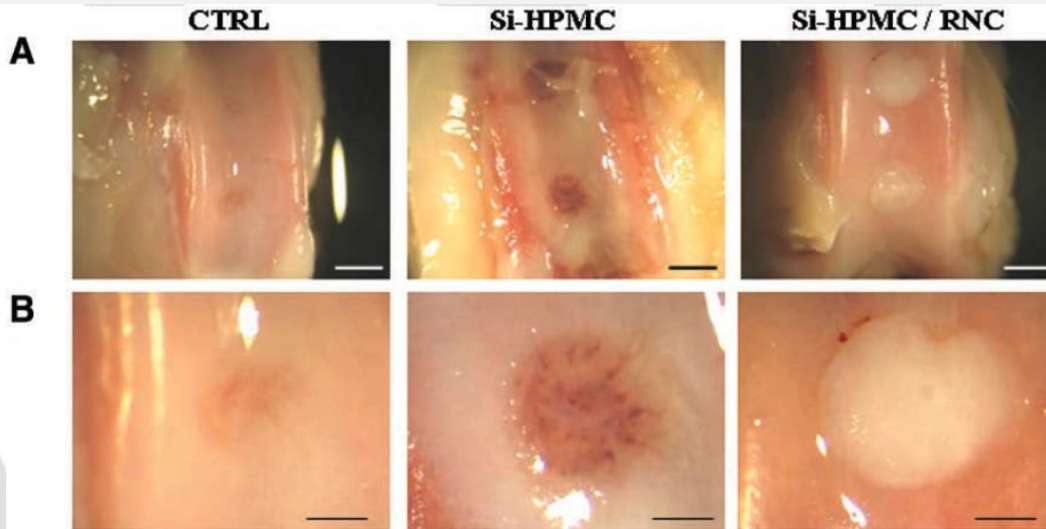
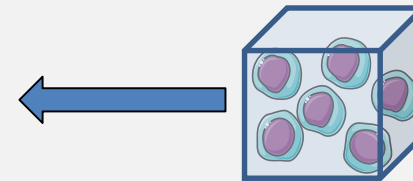
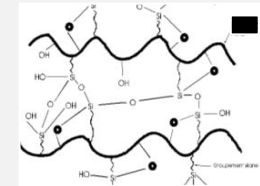
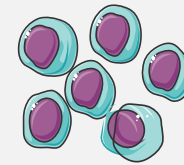
Representation of the self-hardening kinetics of P(6)3% 0.05 M in function of the pH.  
X. Bourges et al. / Advances in Colloid and Interface Science 99 (2002) 215–228

# An Injectable Cellulose-Based Hydrogel for the Transfer of Autologous Nasal Chondrocytes in Articular Cartilage Defects

C. Vinatier,<sup>1,2</sup> O. Gauthier,<sup>1,3</sup> A. Fatimi,<sup>1,3</sup> C. Merceron,<sup>1,3</sup> M. Masson,<sup>1,3</sup> A. Moreau,<sup>4</sup> F. Moreau,<sup>1,3</sup> B. Fellah,<sup>1,3</sup> P. Weiss,<sup>1,3,5</sup> J. Guicheux<sup>1,3</sup>

## For cartilage

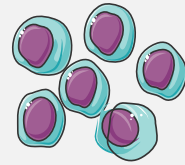
Autologous  
Nasal chondrocytes



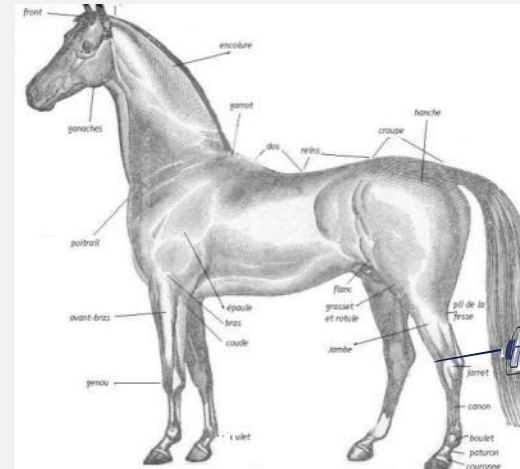
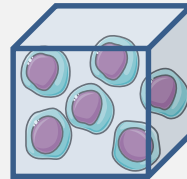
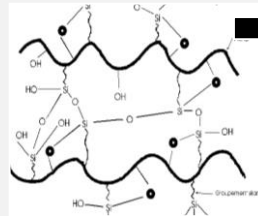




## Cartilage engineering « Proof of concept in large animal and in full-thickness cartilage defect »

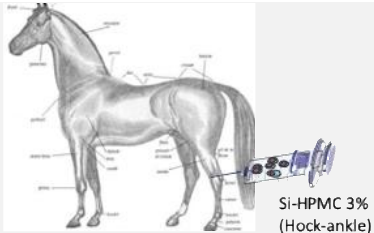


Autologous  
Nasal chondrocytes



Si-HPMC 3%  
(Hock-ankle)

Arthroscopic injection  
(cartilage defect)

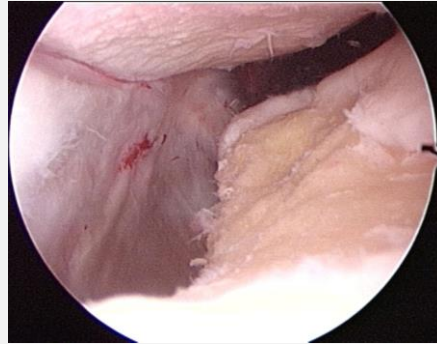


Arthroscopic injection  
(cartilage defect)

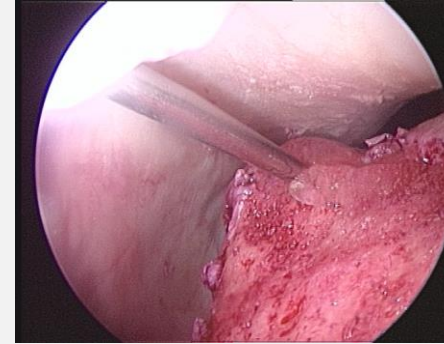
Si-HPMC 3%  
(Hock-ankle)

# Equine clinical case

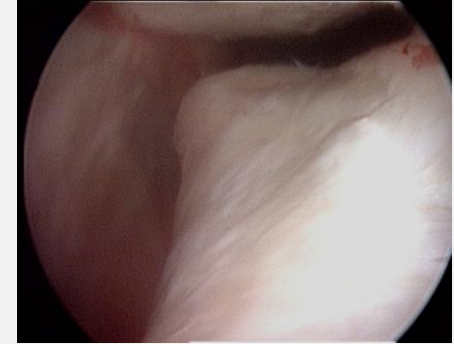
## stifle (Knee)



Cartilage defect



Injection ENC/Si-HPMC



13 weeks

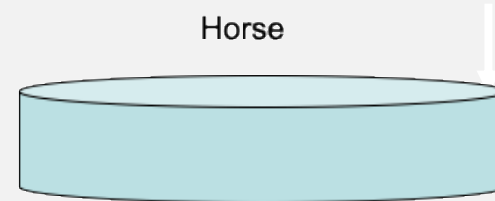


Cartilaginous tissue formation with smooth surface



Rabbit

Height = Diameter



Horse

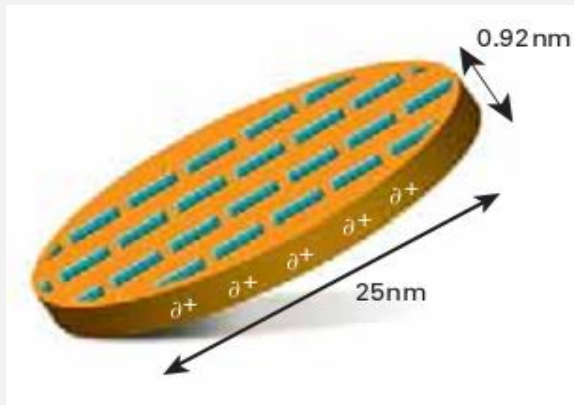
$H < D$

- Hydrogel is weak and does not support unfavourable cavity design
- This Si-HPMC seems bond to the cartilage surface



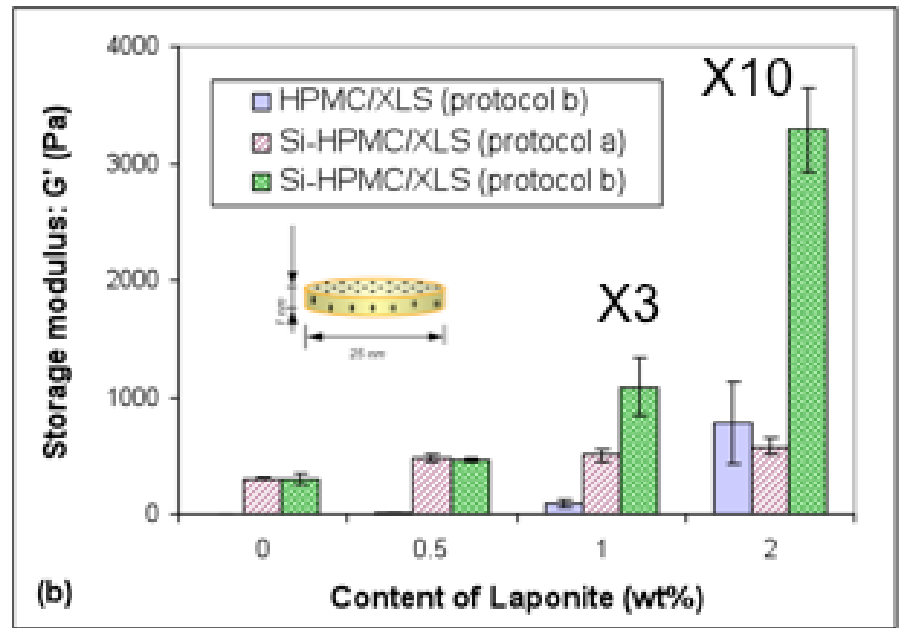
# Laponites (XLS et XLG)

Clay nano disc  
Silicate based  
Diameter : 25 nm  
Thickness : 1 nm

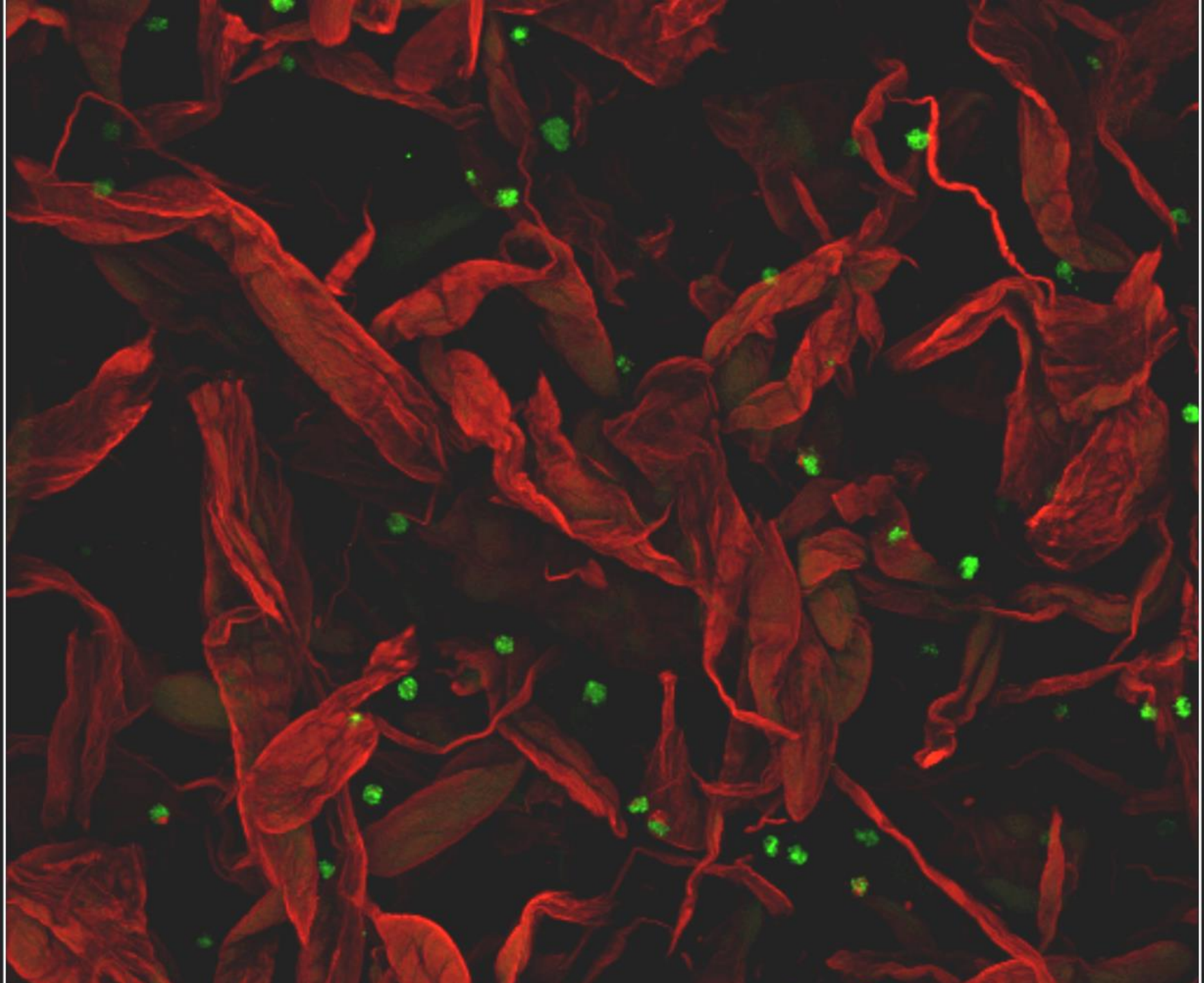


XLS 55% silicate, 26% MgO, 1% LiO, 3% Na<sub>2</sub>O, 4% H<sub>3</sub>PO<sub>4</sub>  
 XLG 60% silicate, 28% MgO, 1% LiO, 3% Na<sub>2</sub>O.

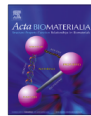
Laponite (silicate clays)











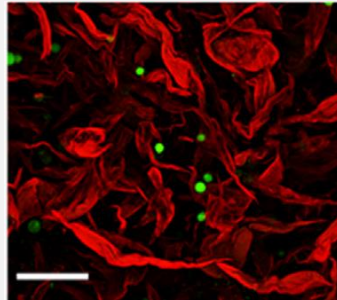
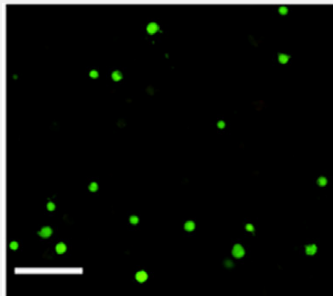
# Institut Thématique Multi-Organismes Technologies pour la santé

Full length article

Laponite nanoparticle-associated silated hydroxypropylmethyl cellulose as an injectable reinforced interpenetrating network hydrogel for cartilage tissue engineering



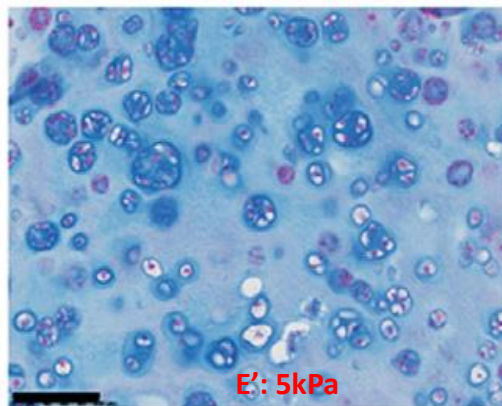
Cécile Boyer<sup>a,b,1</sup>, Lara Figueredo<sup>a,b,1</sup>, Richard Pace<sup>a,b</sup>, Julie Lesoeur<sup>a,b,d</sup>, Thierry Rouillon<sup>a,b</sup>, Catherine Le Visage<sup>a,b</sup>, Jean-François Tassin<sup>c</sup>, Pierre Weiss<sup>a,b,c,\*</sup>, Jerome Guicheux<sup>a,b,c,2</sup>, Gildas Rethore<sup>a,b,c,2</sup>



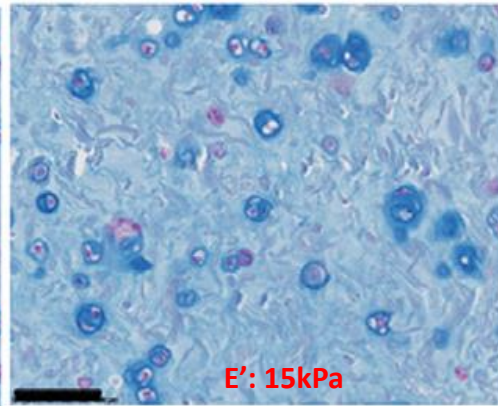
Alcian Blue

0% XLG

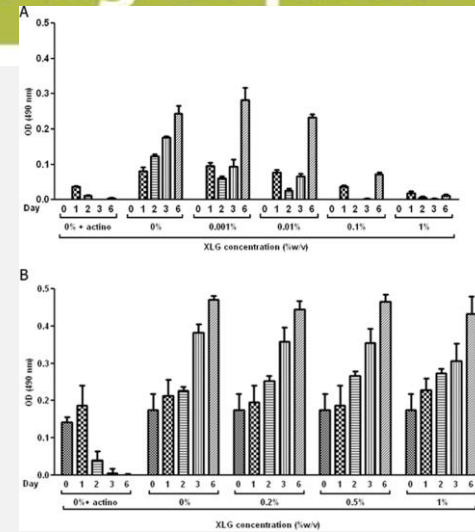
1 % XLG



E': 5kPa

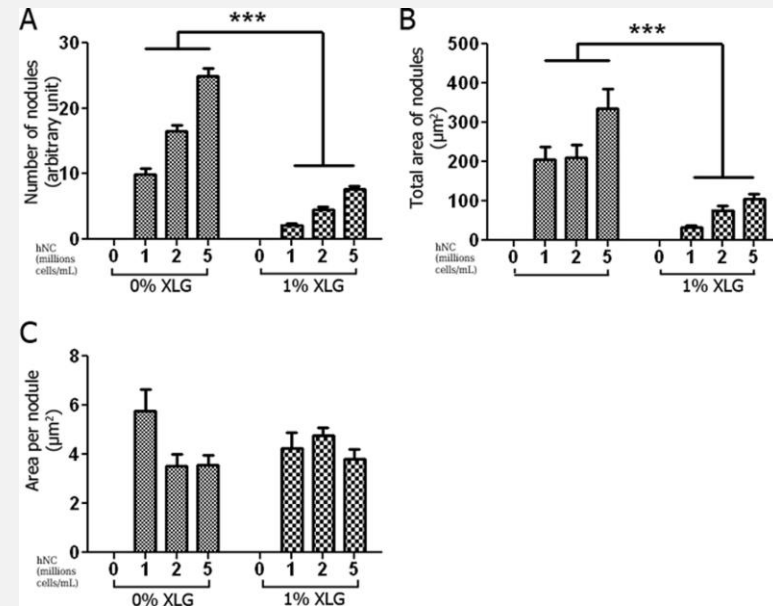


E': 15kPa



2D free laponites

2D with laponite in the hydrogel on the top of the cells

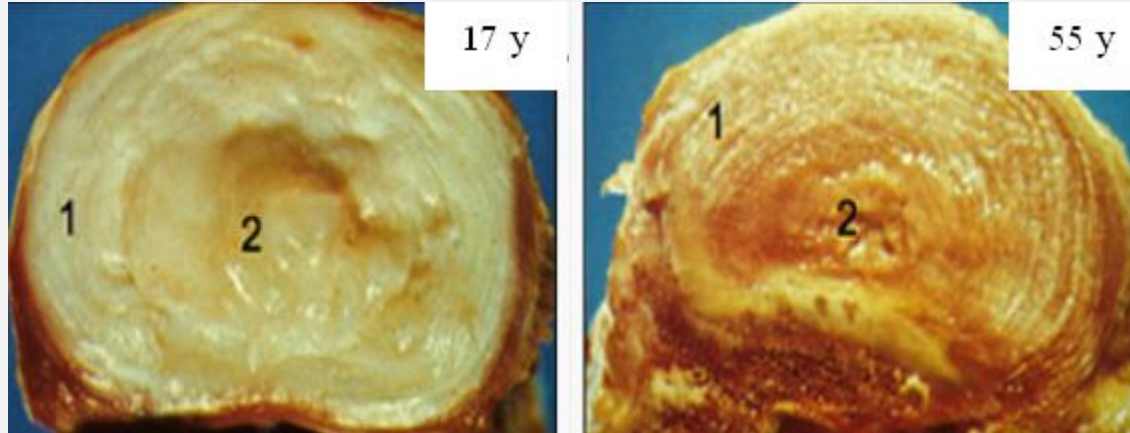
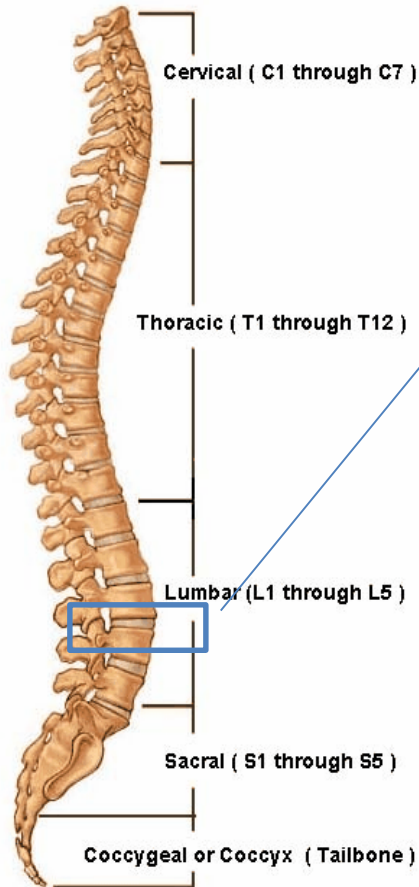


Subcutaneous implantation of hNC with Si-HPMC/XLG hydrogels in a nude mice model

Multi parameters



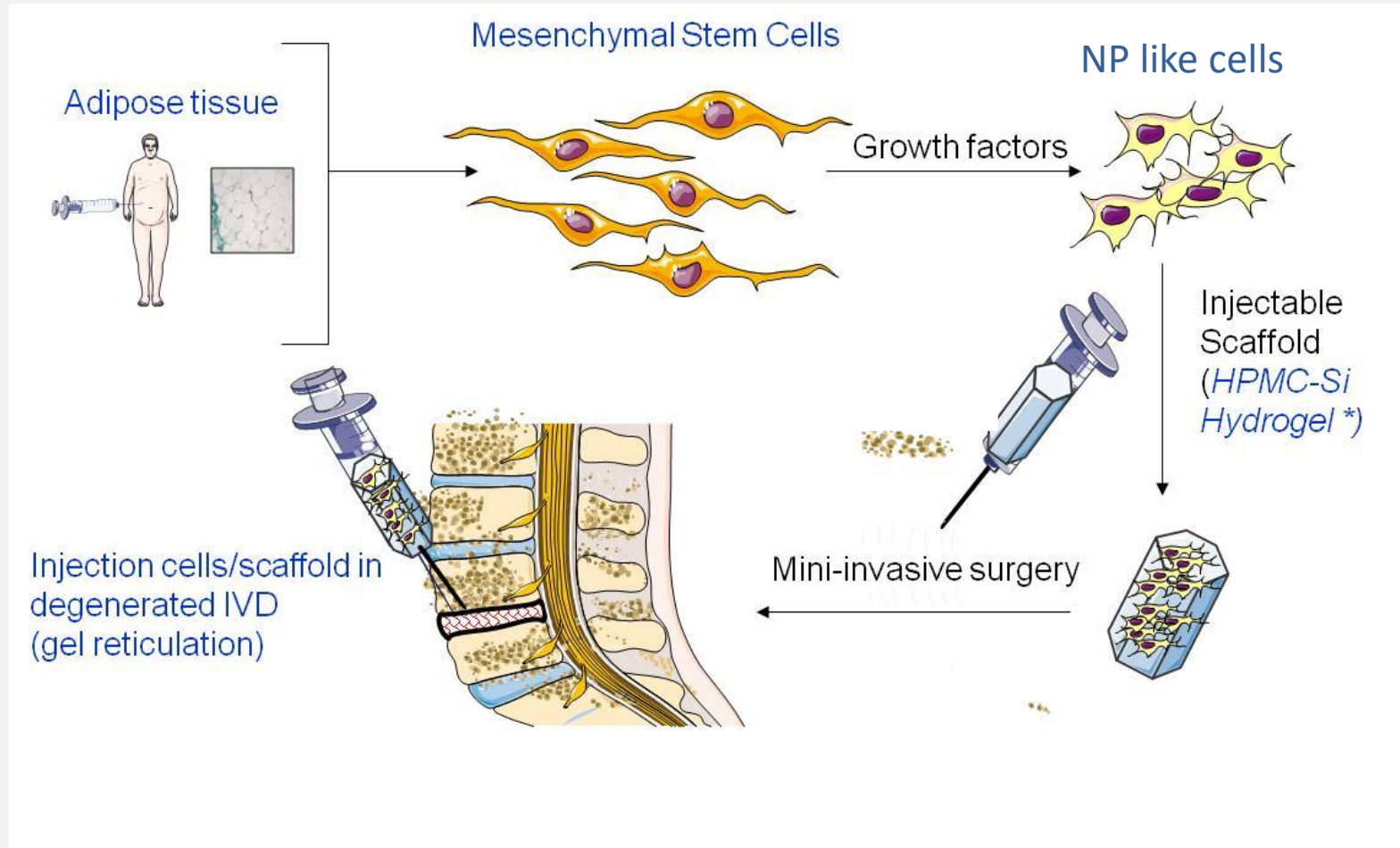
## Intervertebral disc degeneration : IVD



	Healthy	Degenerated
Cell density	+	-
PG amount	+++	+
Hydration	+++	+



## IVD Tissue engineering strategy







# Bone Tissue engineering

## 1) CaP

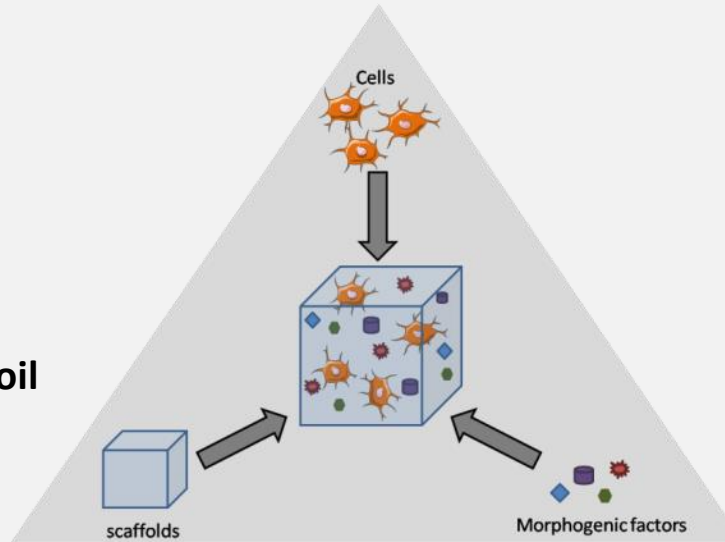


Biomaterials like Soil



## 2) Hydrogels ?

cells like seeds



GFs like fertilizer

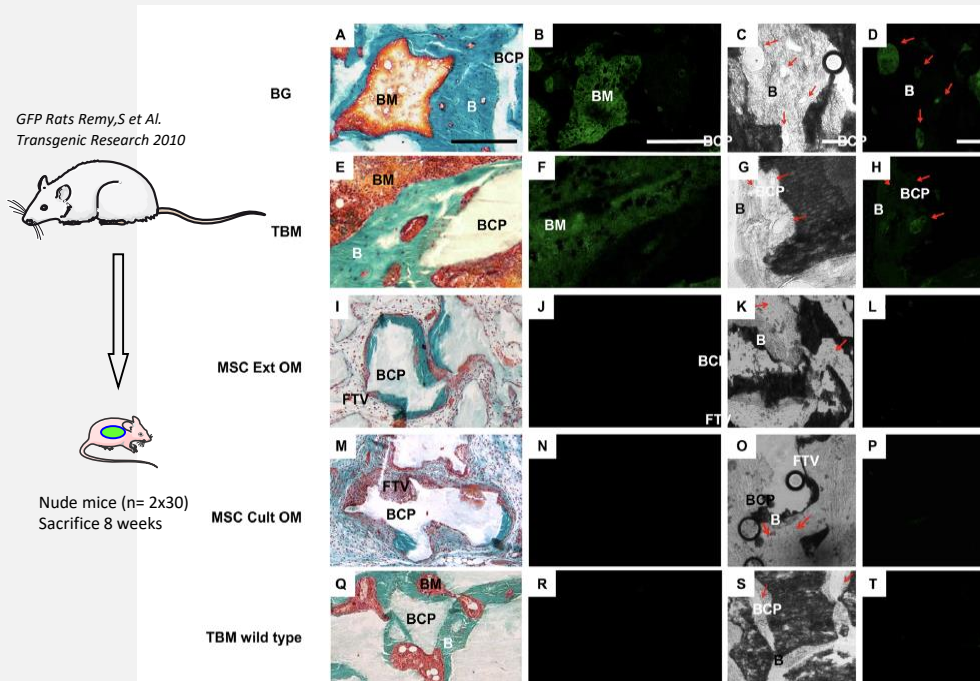




# Determining a Clinically Relevant Strategy for Bone Tissue Engineering: An "All-in-One" Study in Nude Mice

Pierre Corre<sup>1,2,5\*</sup>, Christophe Merceron<sup>1,5\*</sup>, Caroline Vignes<sup>1,5</sup>, Sophie Sourice<sup>1,5</sup>, Martial Masson<sup>1,5</sup>, Nicolas Durand<sup>1,3,5</sup>, Florent Espitalier<sup>1,3,5</sup>, Paul Pilet<sup>1,5</sup>, Thomas Cordonnier<sup>1,5</sup>, Jacques Mercier<sup>2,5</sup>, Séverine Remy<sup>4</sup>, Ignacio Anegon<sup>4</sup>, Pierre Weiss<sup>1,5\*</sup>, Jérôme Guicheux<sup>1,5\*</sup>

## Bone Tissue Engineering using CaP ceramics and BMSC ?



**Figure 7. "In vivo" tracking of donor cells.** Goldner trichrome staining (A, E, I, M and Q). B: bone, BCP: biphasic calcium phosphate, BM: bone marrow, FVT: fibrovascular tissue. Bar: 250  $\mu$ m. Green fluorescence of GFP retrieved in subcutaneous implants (B, F, J, N and R). Nude mice implanted with non-GFP BM were used as negative controls (TBM wild type). Bar: 250  $\mu$ m. Transmitted light showing vessels in connective tissues surrounding the BCP granules or in newly formed bone (red arrow) (C, G, K, O and S) Bar: 100  $\mu$ m. Fluorescent light showing vessels only in TBM and BG groups (red arrow) (D, H, L, P and T) Bar: 100  $\mu$ m.  
doi:10.1371/journal.pone.0081599.g007

## MSCs fate ?

→ Cells die after 4 weeks of implantation in bone

*J. Cell. Mol. Med. Vol 15, No 7, 2011 pp. 1505-1514*

**Survival and function of mesenchymal stem cells (MSCs) depend on glucose to overcome exposure to long-term, severe and continuous hypoxia**

M. Descheppe<sup>1</sup>, K. Oudina<sup>1</sup>, B. David<sup>1,2</sup>, V. Myrtil<sup>1</sup>, C. Collet<sup>1</sup>, M. Bensidhoum<sup>1</sup>, D. Logeart-Avramoglou<sup>1</sup>, H. Petite<sup>1,3</sup>

→ Hypoxia with Glucose doesn't kill MSC : Ischemia is the problem

nature  
medicine

Mesenchymal stem cell-based tissue regeneration is governed by recipient T lymphocytes via IFN- $\gamma$  and TNF- $\alpha$

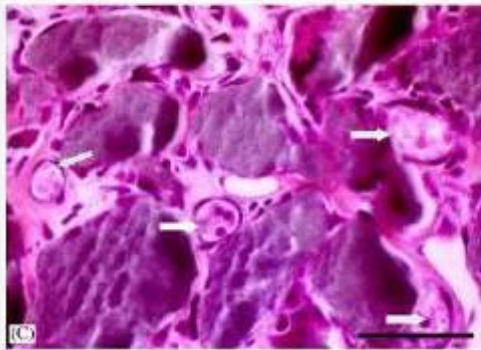
Yi Liu<sup>1,2</sup>, Lei Wang<sup>1,3</sup>, Takashi Kikui<sup>1</sup>, Kentaro Akiyama<sup>1</sup>, Chider Chen<sup>1</sup>, Xingtian Xu<sup>1,4</sup>, Ruili Yang<sup>1</sup>, WanJun Chen<sup>1</sup>, Songlin Wang<sup>2</sup> & Songtao Shi<sup>1</sup>

→ T cells regulate autologous MSC

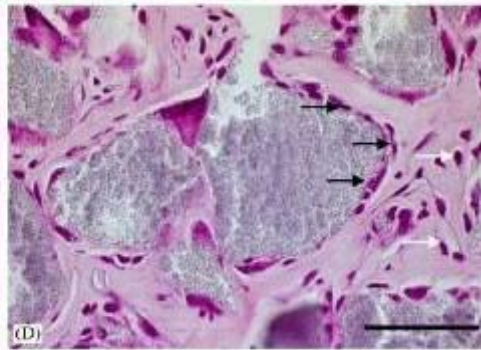


# Bone Tissue reconstruction *in vivo* with an hydrogel

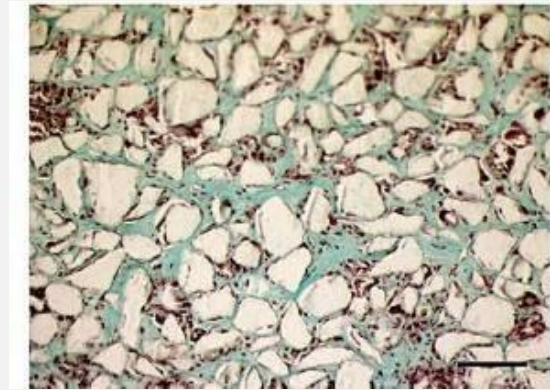
BMSC with Si-HPMC / BCP formulation implanted under skin of mice for 4 weeks



Blood vessels



Osteoblasts and osteocytes



Goldner staining paraffin sections



Available online at [www.sciencedirect.com](http://www.sciencedirect.com)

SCIENCE @ DIRECT®

Biomedicine 37 (2006) 1256–1264

Blomaterials

[www.elsevier.com/locate/blomaterials](http://www.elsevier.com/locate/blomaterials)

Ectopic bone formation using an injectable biphasic calcium phosphate/Si-HPMC hydrogel composite loaded with undifferentiated bone marrow stromal cells

Christophe Trojani<sup>a,b</sup>, Florian Boukhechba<sup>a</sup>, Jean-Claude Seimeca<sup>a</sup>, Fanny Vandenbos<sup>c</sup>, Jean-François Michiels<sup>d</sup>, Guy Duculsi<sup>d</sup>, Pascal Boileau<sup>b</sup>, Pierre Weiss<sup>d</sup>, Georges F. Carle<sup>a</sup>, Nathalie Rochet<sup>b,e,\*</sup>

Ectopic bone formation was available in mice model using MSC in Hydrogel  
 → enough nutrients diffusion  
 → Hydrogel is a shield against immune system ?



# The futur

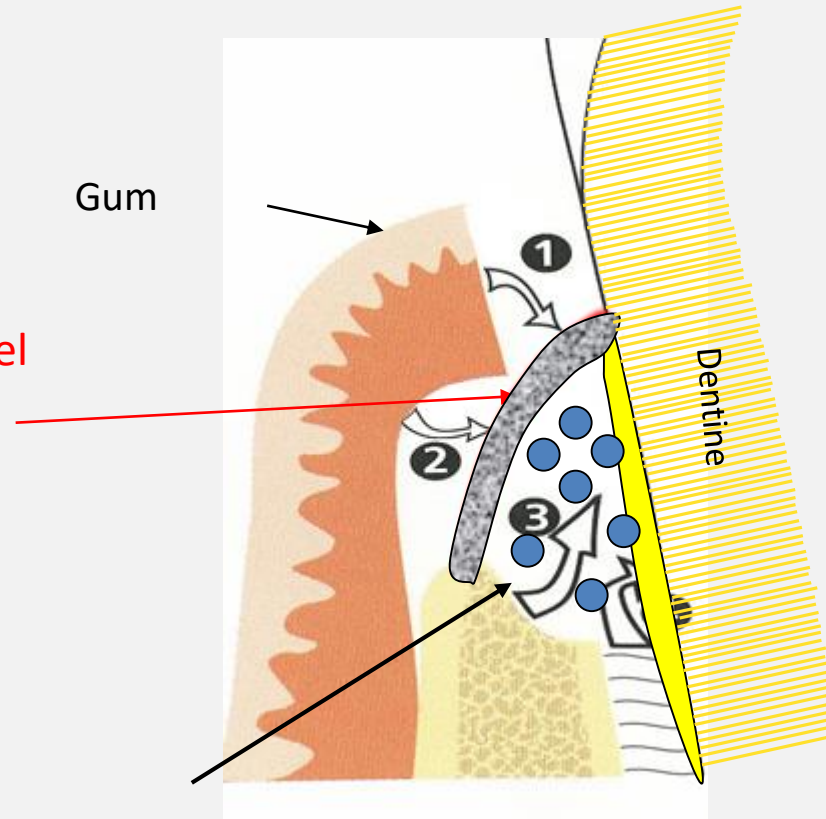


# GTR using stratification



Periodontal disease

Injectable Hydrogel  
Membrane



- Easy and quick protocol
- Spatial control of tissue differentiation

IBS : Injectable bone substitute = fast bone ingrowth





Pauline Chichirico  
(Posture program)

# In situ photochemical crosslinking of hydrogel membrane for Guided Tissue Regeneration

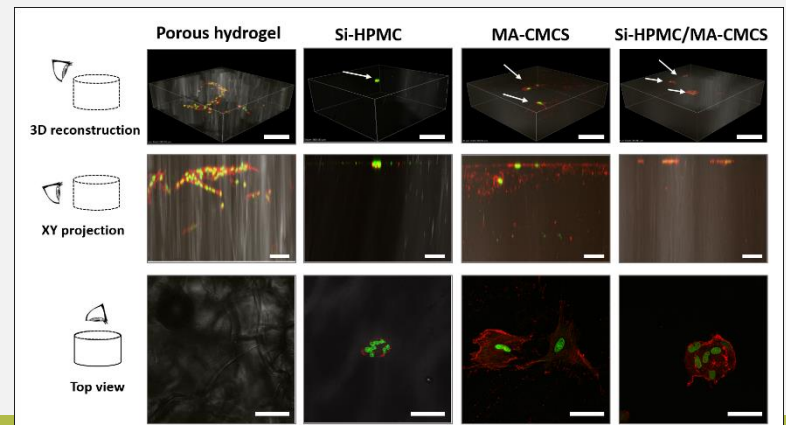
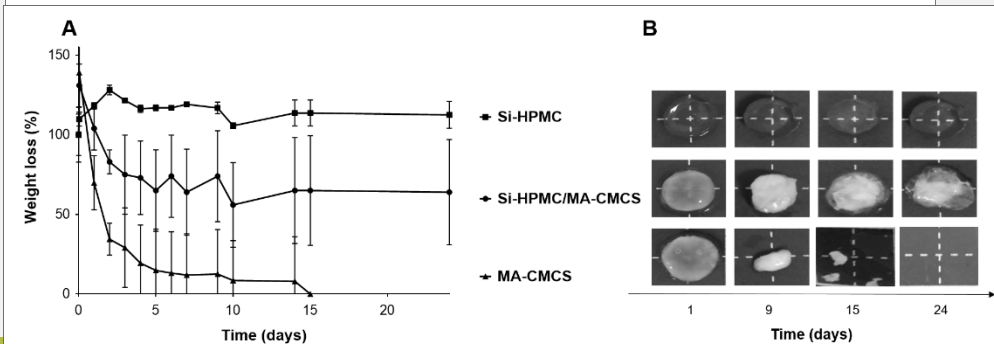
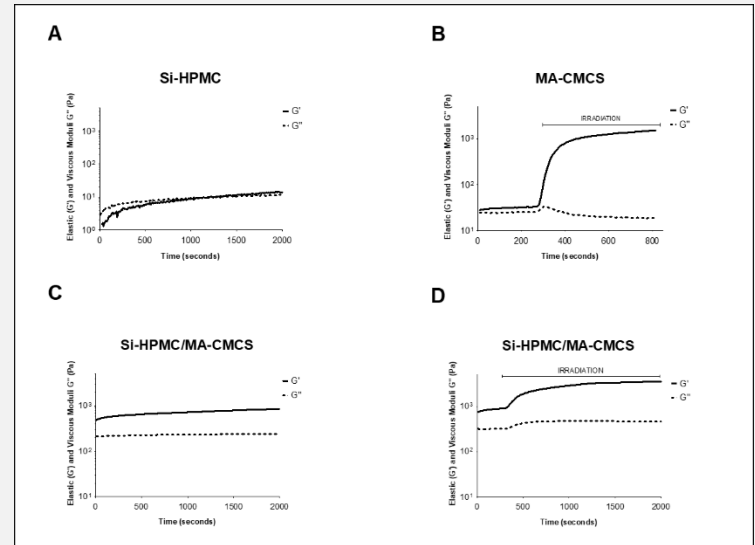
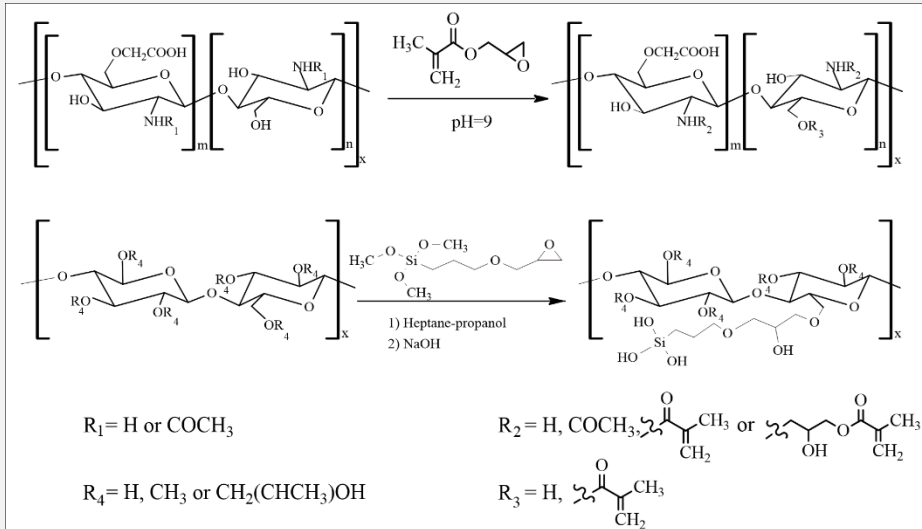
Pauline Marie Chichirico <sup>a, b</sup>, Raphael Riva <sup>a</sup>, Jean-Michel Thomassin <sup>a</sup>, Julie Lesueur <sup>b, c, d</sup>, Xavier Struillou <sup>b, c, e</sup>, Catherine Le Visage <sup>b, c</sup>, Christine Jérôme <sup>a, 1</sup>, Pierre Weiss <sup>b, c, e, 1</sup> ✉

Show more

<https://doi.org/10.1016/j.dental.2018.09.017>

Get rights and content

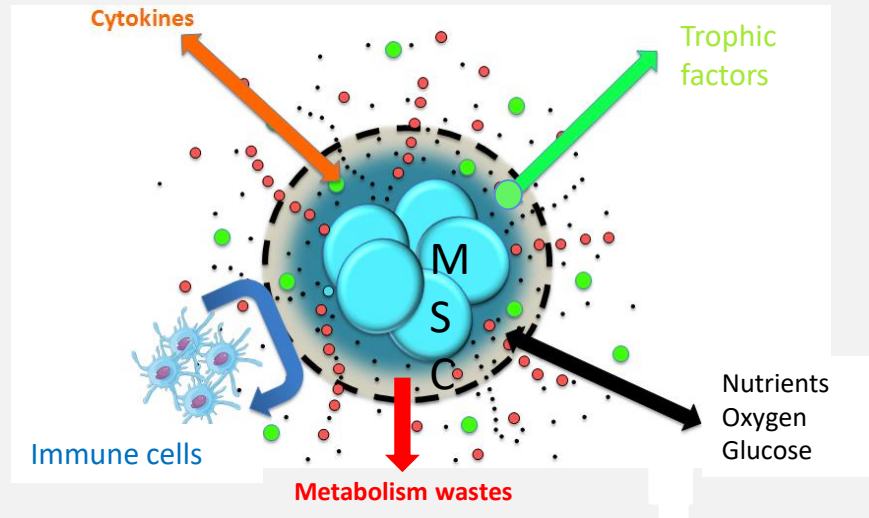
# GTR membrane





### Assisted cell therapy

#### Embedded MSC in hydrogel



Hindawi  
Stem Cells International  
Volume 2017, Article ID 9003598, 11 pages  
<https://doi.org/10.1155/2017/9003598>



#### Research Article

**Polysaccharide Hydrogels Support the Long-Term Viability of Encapsulated Human Mesenchymal Stem Cells and Their Ability to Secrete Immunomodulatory Factors**

Fahd Hached,<sup>1,2</sup> Claire Vinatier,<sup>1,3</sup> Pierre-Gabriel Pinta,<sup>1,4</sup> Philippe Hulín,<sup>5</sup>  
Catherine Le Visage,<sup>1,3</sup> Pierre Weiss,<sup>1,3,6</sup> Jérôme Guicheux,<sup>1,3,6</sup>  
Aurélien Billon-Chabaud,<sup>1,2</sup> and Gaël Grimandi<sup>1,2,4</sup>



→ « IXBONE » ANR Program  
→ LFC Bioregate program



# Conclusion

- Regenerative medicine / Innovation
  - >10 SMEs
  - >10 Medical applications
- New biomaterials / Closer to ECM
- 3D printing and Bio printing
  - ART Bordeaux, ECN...
  - Personalized medicine
  - Bio-Ink
- Assisted cell therapy
- Extra cellular vesicles

THANK  
YOU



- Pierre Weiss

INSERM U 1229

**RMES Regenerative medicine and skeleton,**

University of Nantes, 1 place Alexis Ricordeau, 44042 Nantes, France.

E-mail : [pierre.weiss@univ-nantes.fr](mailto:pierre.weiss@univ-nantes.fr)

**Resercher ID : P-1372-2014**