

21 Avril 2010



Multi-modalities and Non-invasive Imaging In Tissue Engineering : From Microscopy to Macroscopy

PTIBC-IBISA

Dominique Dumas

S. Hupont, JF. Stoltz, P. Gillet

Université Henri Poincaré Nancy I

UMR 7561 CNRS - Directeur : Jacques Magdalou

FR3209 CNRS . Directeur : Patrick Netter

GDR2588CNRS (L. Héliot)/RTfmf (D.Dumas)

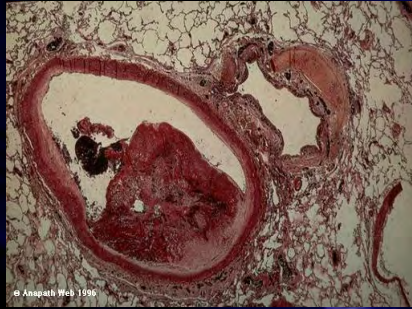
ITMO BCBD - A. Le Bivic

ITMO TS - J. Grassi



• **3D Imaging : Medical Tissue Engineering (stem cells)**

**Endothelial Cells
Blood vessel**



**Occlusion
Atherosclerosis**

**Hyalin chondrocytes
Cartilage matrix**



**Degenerative disease
Osteoarthritis**

Ligament fibroblasts



LCA Rupture

Thick and opaque specimens

1998 : 3D deconvolution imaging

2002 : CLSM Confocal Laser Scanning Microscopy

2003 : TPE Multiphoton Microscopy (FLIM)

2006 : FCS

2007 : SHG Microscopy

2009 : MP-SHG Macroscopy (prototype)

2010 : Phase Imaging

Future ...

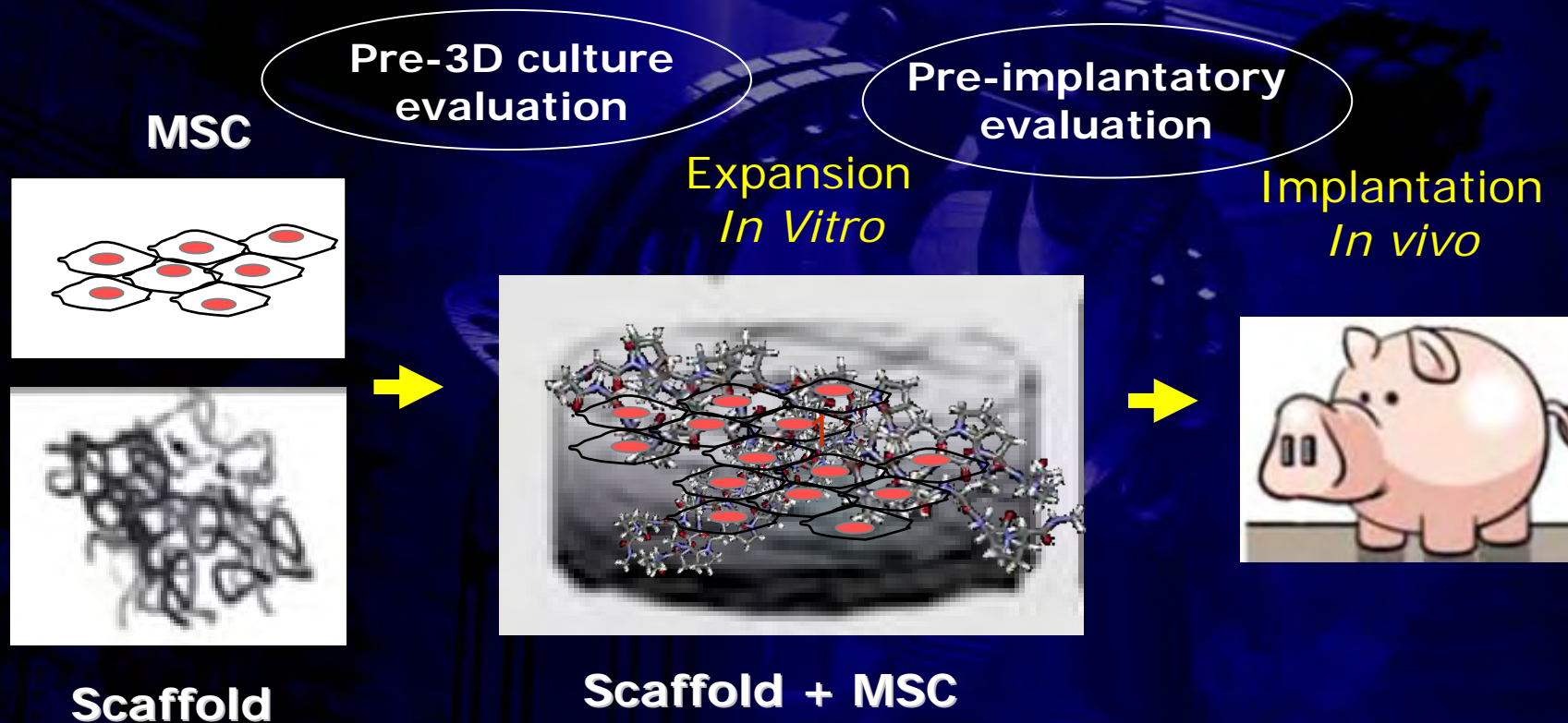
**Core Facility
Service**

Industrial Partnership

**R&D
Collagen imaging
Phase imaging**

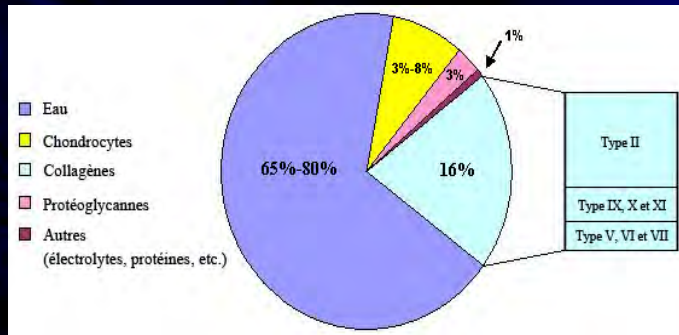
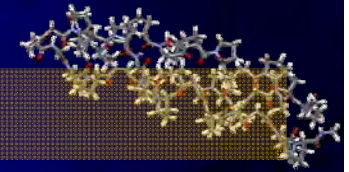
• Collagen for medical scaffold : **Invasiveness 3D network Imaging**

Noninvasive evaluation of MSC –b scaffold for Medical application

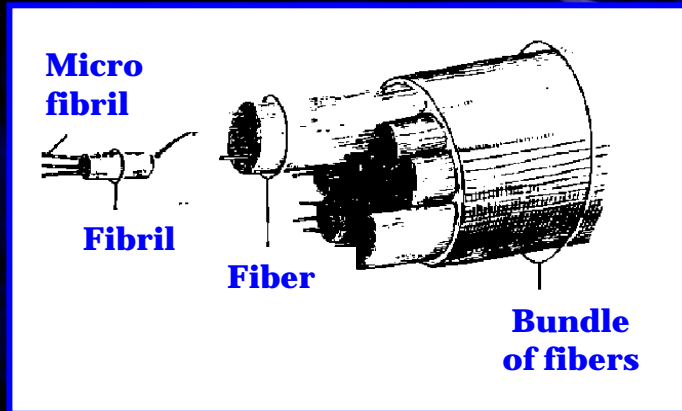


Multiscaled device (from cell to embedded scaffold)
No contrast agent
Qualitative evaluation of **collagen network**
Quantitative evaluation : **cell proliferation / synthesis**

• 3D network : collagen functional



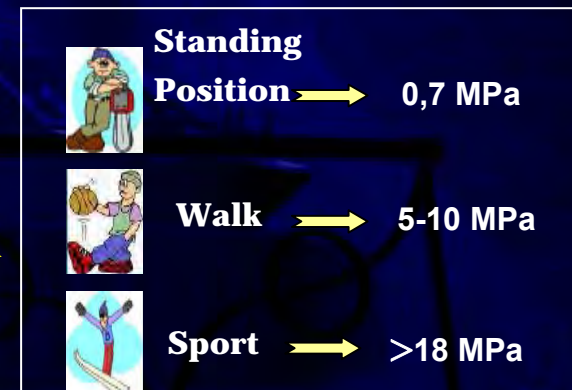
Cartilage Composition



Organization of collagen fibrils into fibers and bundles

	Type	Polymerized form	Representation
Fibrils	I	fibrils	Bone, skin, tendon, ligament, cornea, internal organs, fibrous cartilage
	II	fibrils	cartilage (elastic and hyaline)
	III	fibrils	hypoderm, vessel, hair
	V	fibrils (with type I)	see type I
	XI	fibrils (with type II)	see type II
Associated with fibrils	IX	Lateral connection with type II fibrils	cartilage
	XII	Lateral connection with type II fibrils	Some other tissues
Network	IV	network	Basal lamina
	VII	Anchoring fibrils in basal membranes	epidermis

Evaluation of Functional network Mechanical properties



Interaction light / Matter : IR light for Tissue Imaging

Tissue imaging :

- Cartilage
- Tumors
- Arteries
- Tendon...



Thick and opaque specimens

Advantages of multiphoton (IR) excitation

- Less absorption by biological specimens
→ Deeper penetration (than UV-Visible)
- Less photodamage
→ confinement of excitation to the focal plane

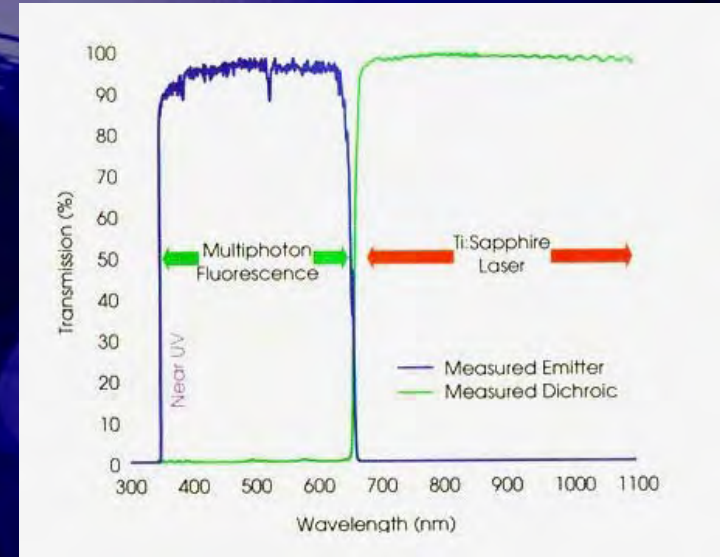
Principle of multiphoton excitation

- Spatio-temporal confinement of photons
- Conditions obtained with short impulsions at high frequency

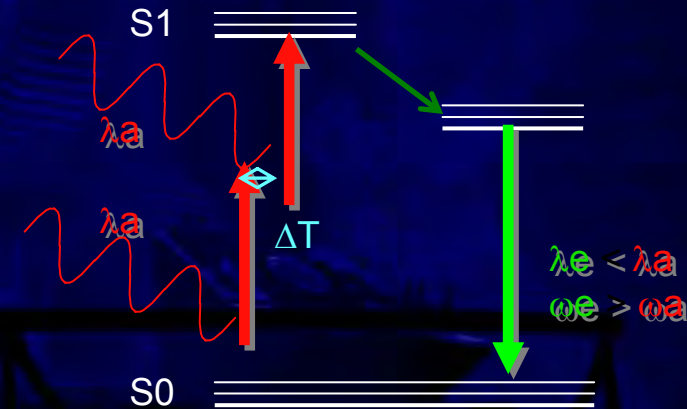


Über Elementarakte mit zwei Quantensprüngen
 Von Maria Göppert-Mayer
 (Göttinger Dissertation)
 (Mit 5 Figuren)
 Einleitung
 Der erste Teil dieser Arbeit beschäftigt sich mit dem
 Zusatzschwächen zweier Lichtquanten in einem Elementarakt

AIP Niels Bohr Library

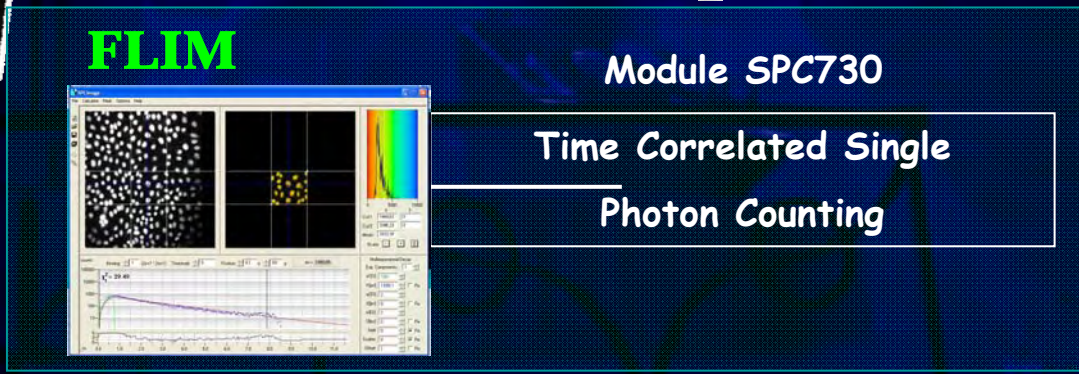
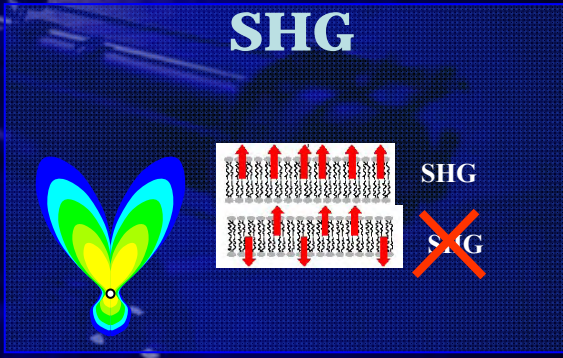
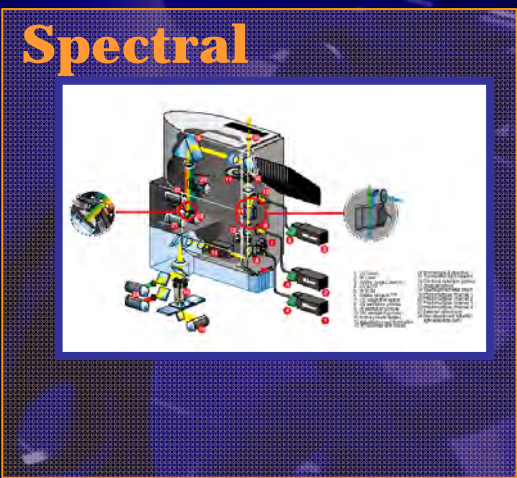
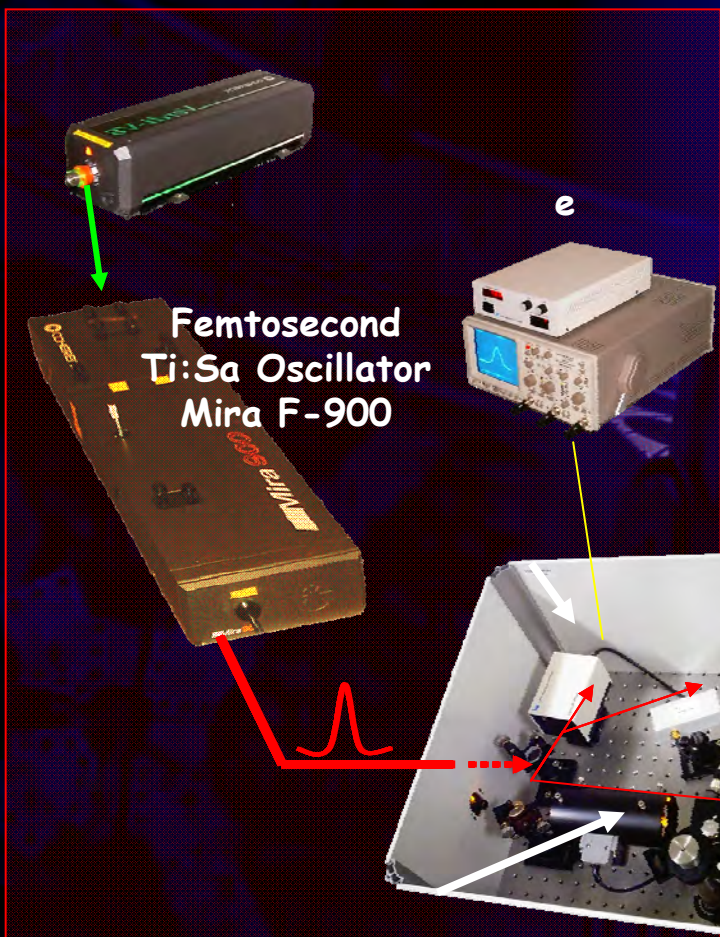


TPE (Two Photon Excitation)



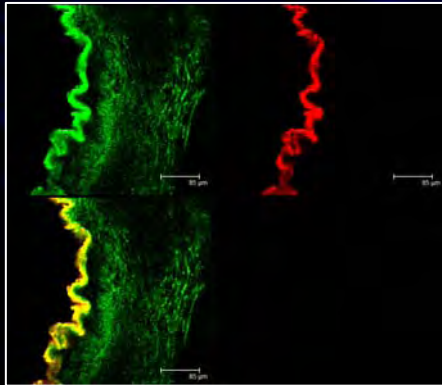
Non linear Absorption
Emission in visible

• IR pulsed laser : modalities imaging techniques

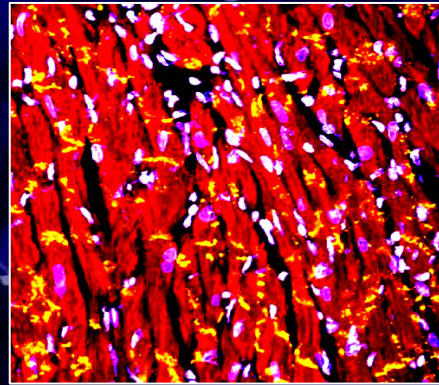


- MP imaging applied to tissue : **fluorescent probe**

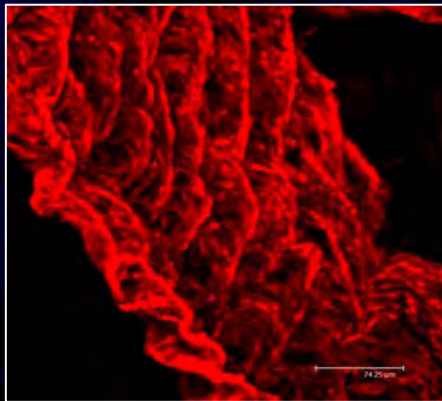
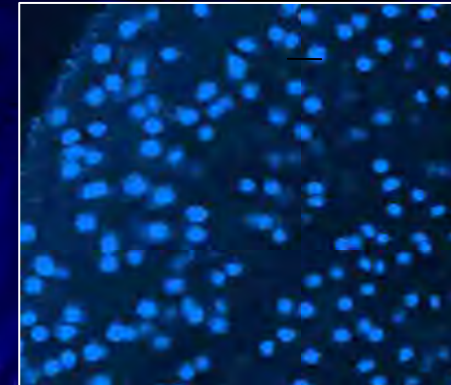
Femoral artery



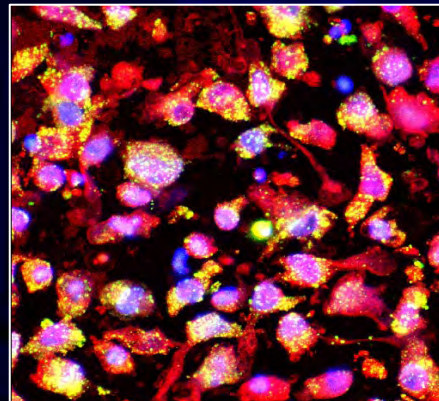
Graft Cardiomyocyte



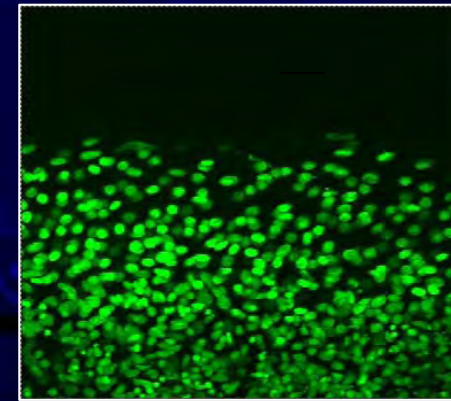
Rat articular cartilage
Chondrocyte (nucleus in blue)
Depth : 219 μm



Femoral artery
+ film PAH- Rhodamine

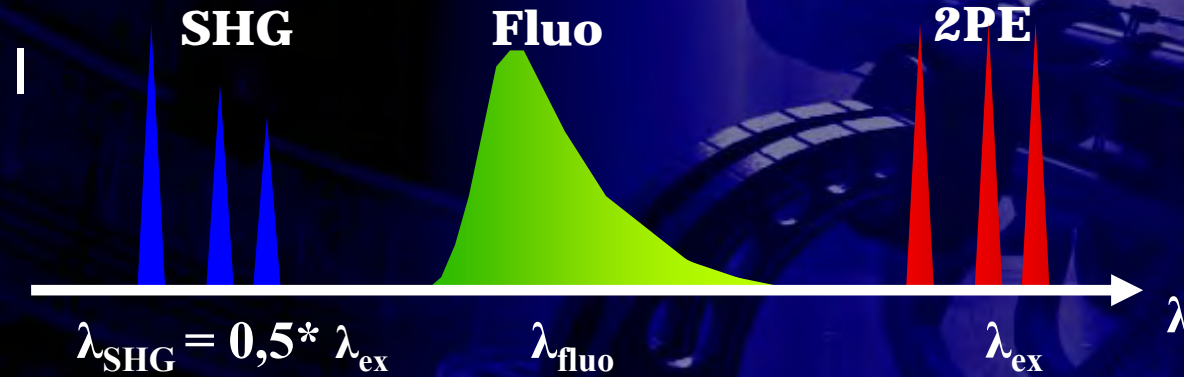


Apoptosis (live/dead)

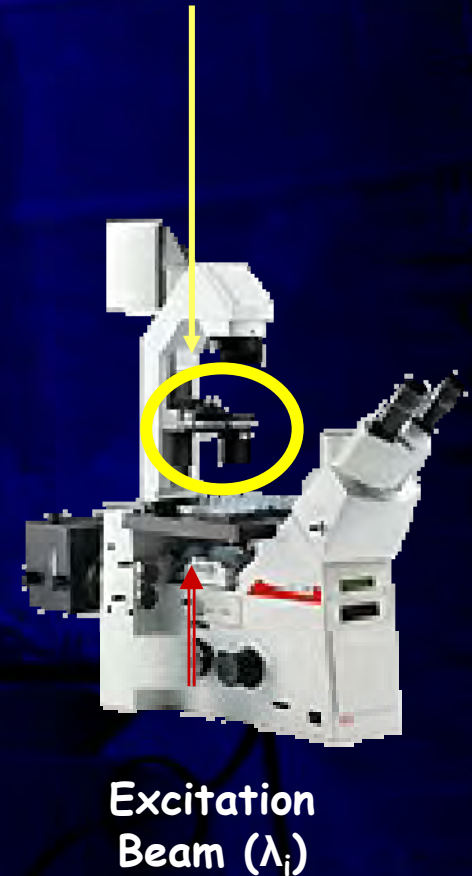
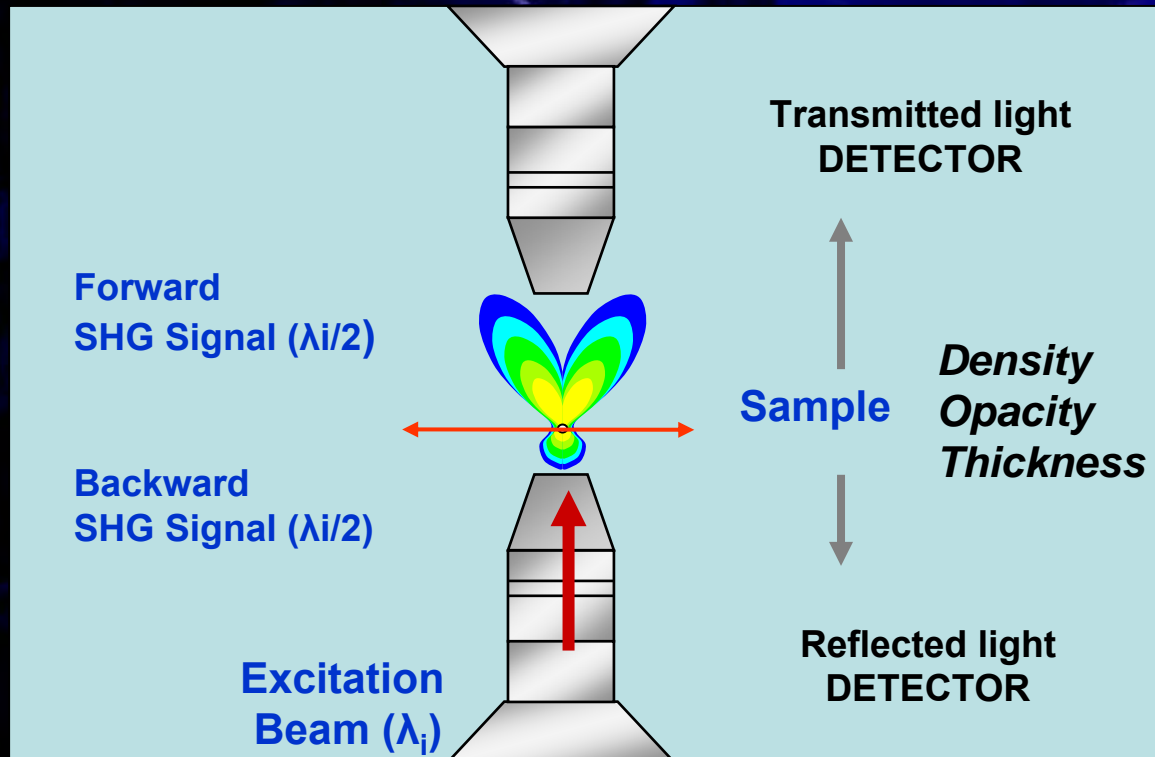


Chondrocyte in alginate bead
Depth : 1600 μm

- SHG : Second Harmonic Generation

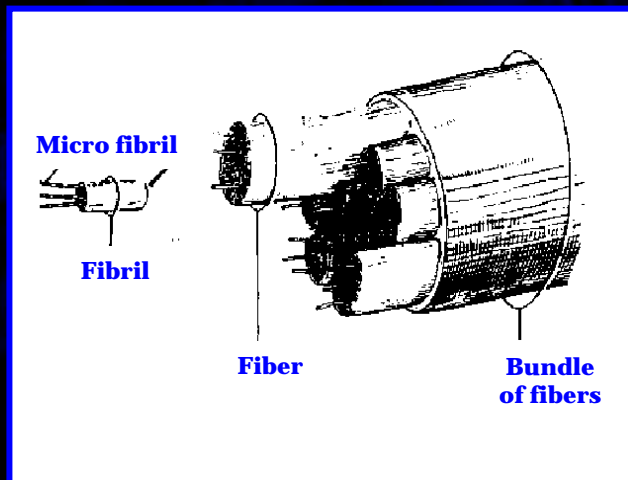


PMT (Transmitted Light)
 BP Filter 400 ± 25 nm
 (=800/2)
 LP Filter 700 nm (IR)

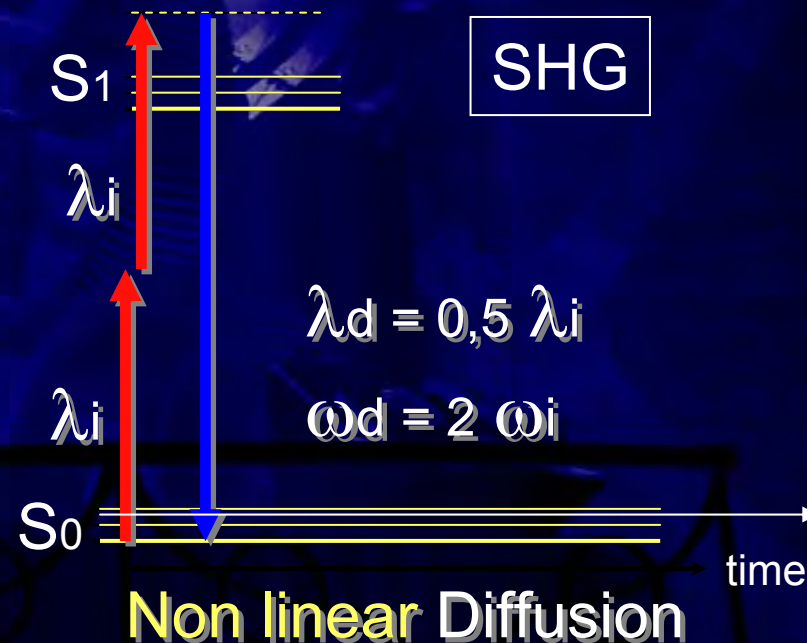


- **SHG collagen : high non linear susceptibility**

- Main tissular component giving rise to SHG : Collagen
- Importance of orientation and arrangement of molecules
- Quadratic dependence on number of molecules
- Quasi instantaneous Generation (fs) → Coherent Signal
- No exogenous dye → Diminution of cytotoxic and phototoxic effects
- No absorption process, no photobleaching (≠ Fluorescence)
- Quadratic dependence on excitation power

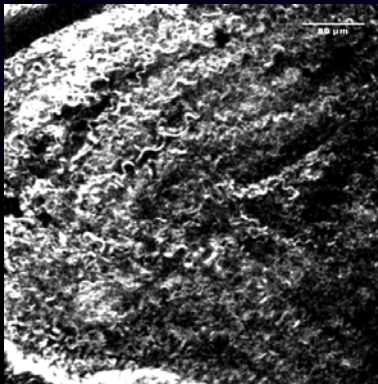


- Crystalline triple-helical structure (left)
- Non-centro symmetric structure

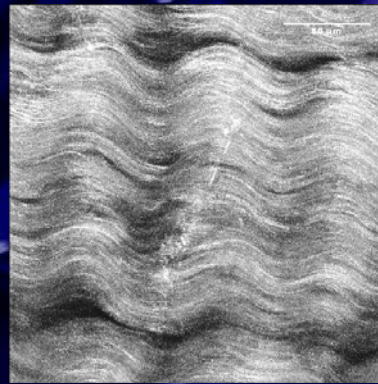


- **SHG : 3D network for functional collagen**

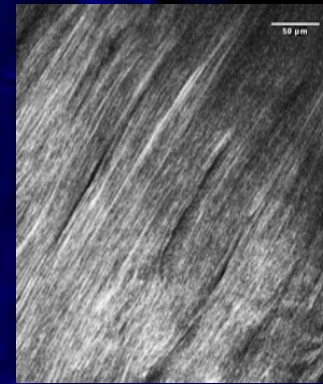
Skin mouse



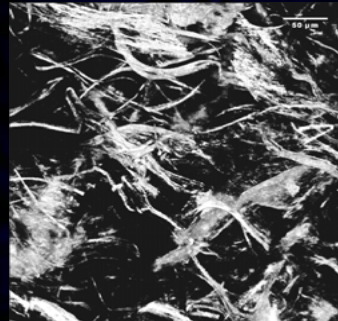
Rabbit Tendon



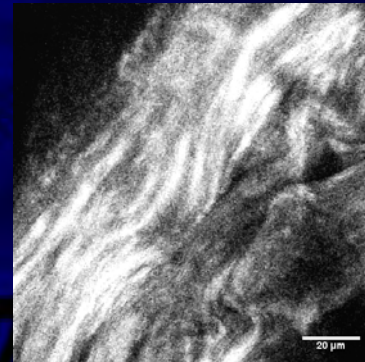
Rabbit muscle



Collagen I
sponge

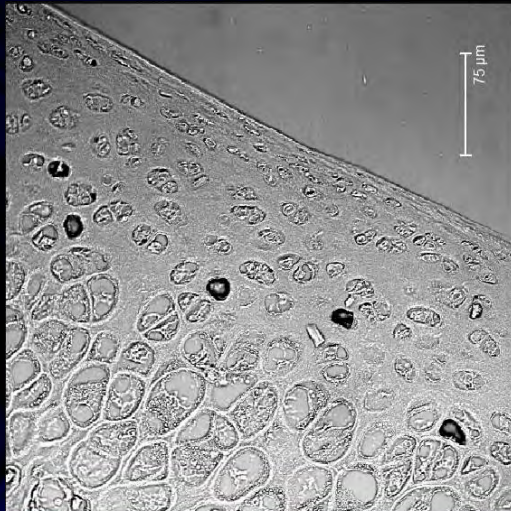


Inter-costal
cartilage
(sternum)

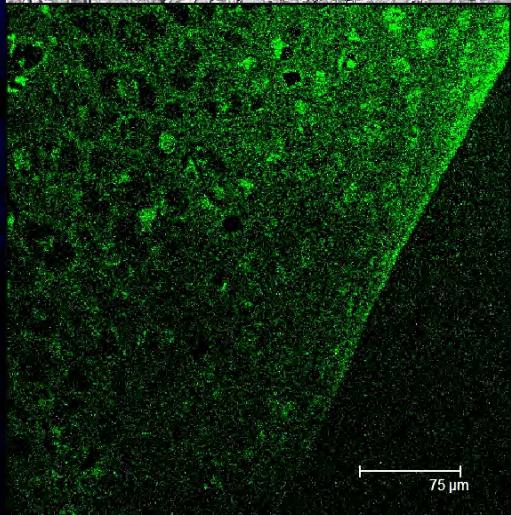


• Cartilage SHG imaging : **quantitative evaluation**

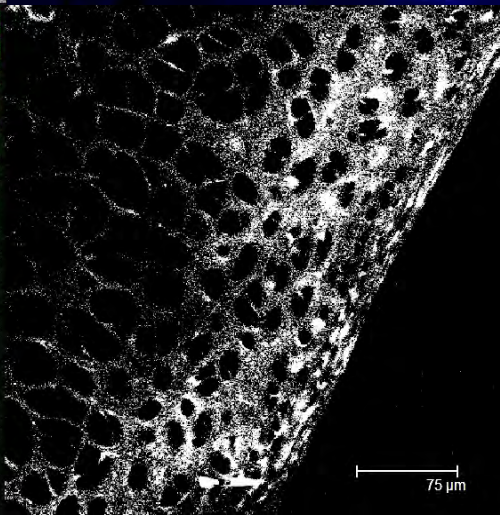
Rat Articular Cartilage



LT



AF



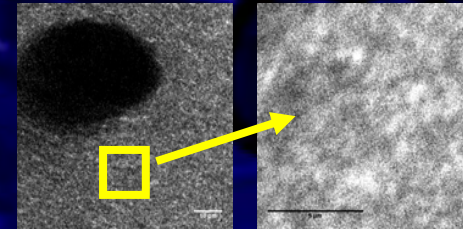
SHG

Textural analysis ?

3D Organisation ?

Denaturation
Degradation
Synthesis

Haralick's analysis



Gray levels : SHG - Collagen

Haralick :

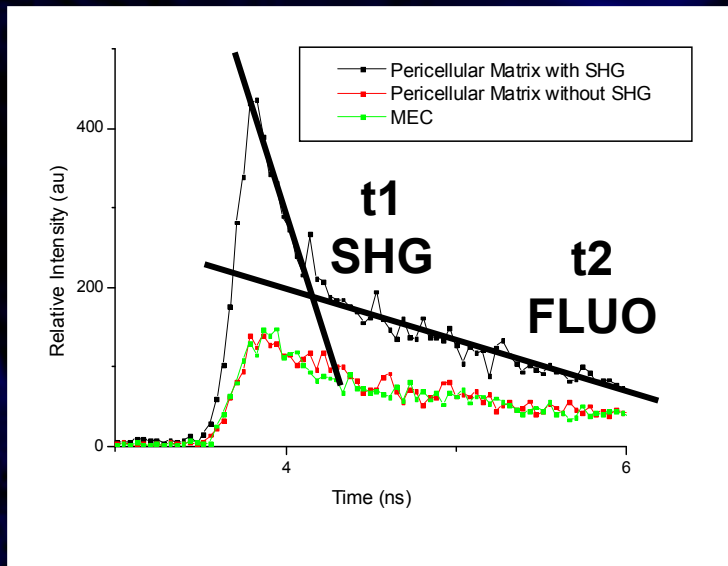
Textural analysis based on cooccurrence matrix

9 parameters of textural elements:

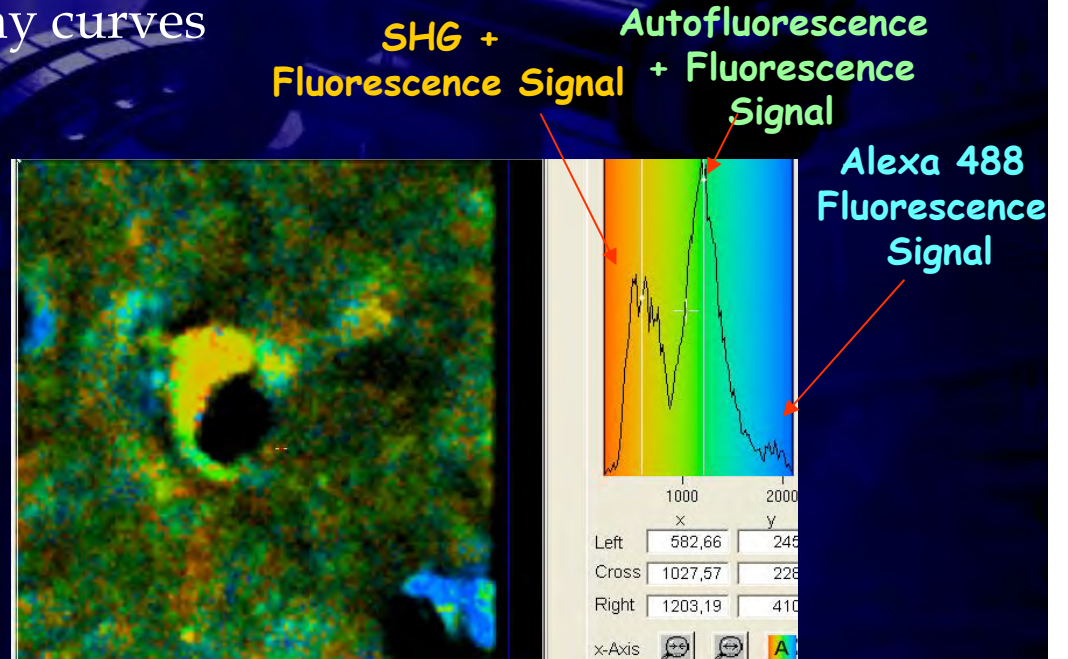
- Homogeneity
- Size
- Linearity
- Contrast
- Variance
- Second Angular Moment ...

- **Micro- TCSPC- SHG imaging : time decay in matrix**

Fluorescence Lifetime Imaging Microscopy
 Biexponential adjustment of Decay curves



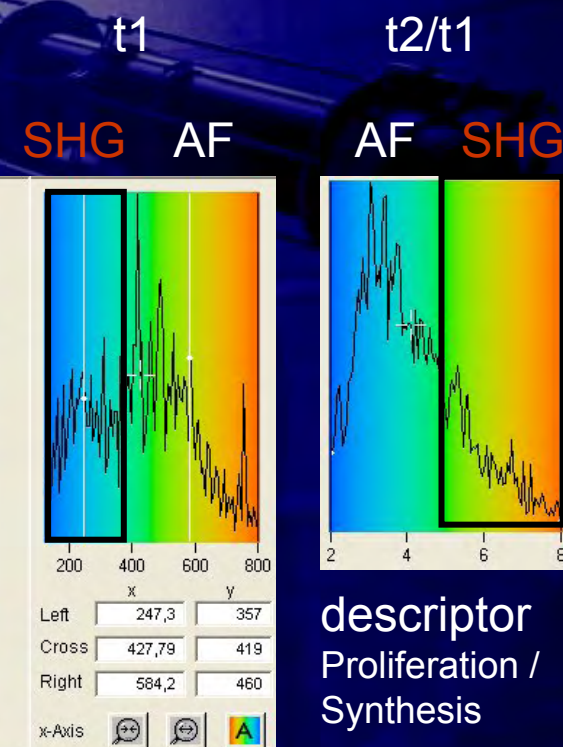
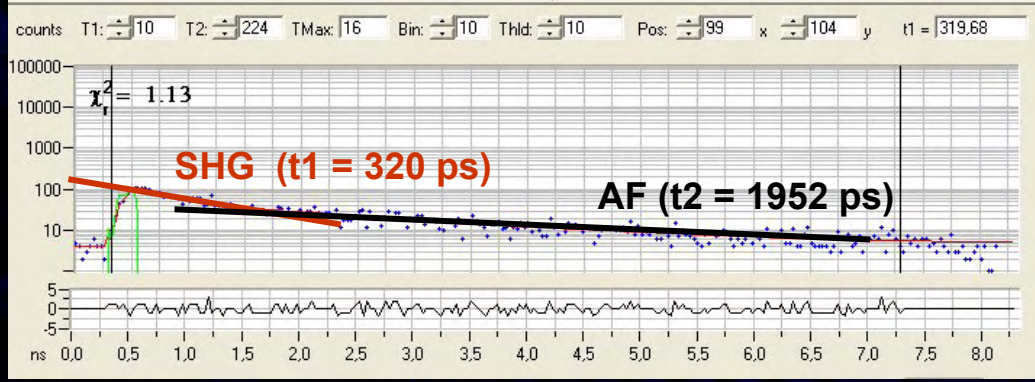
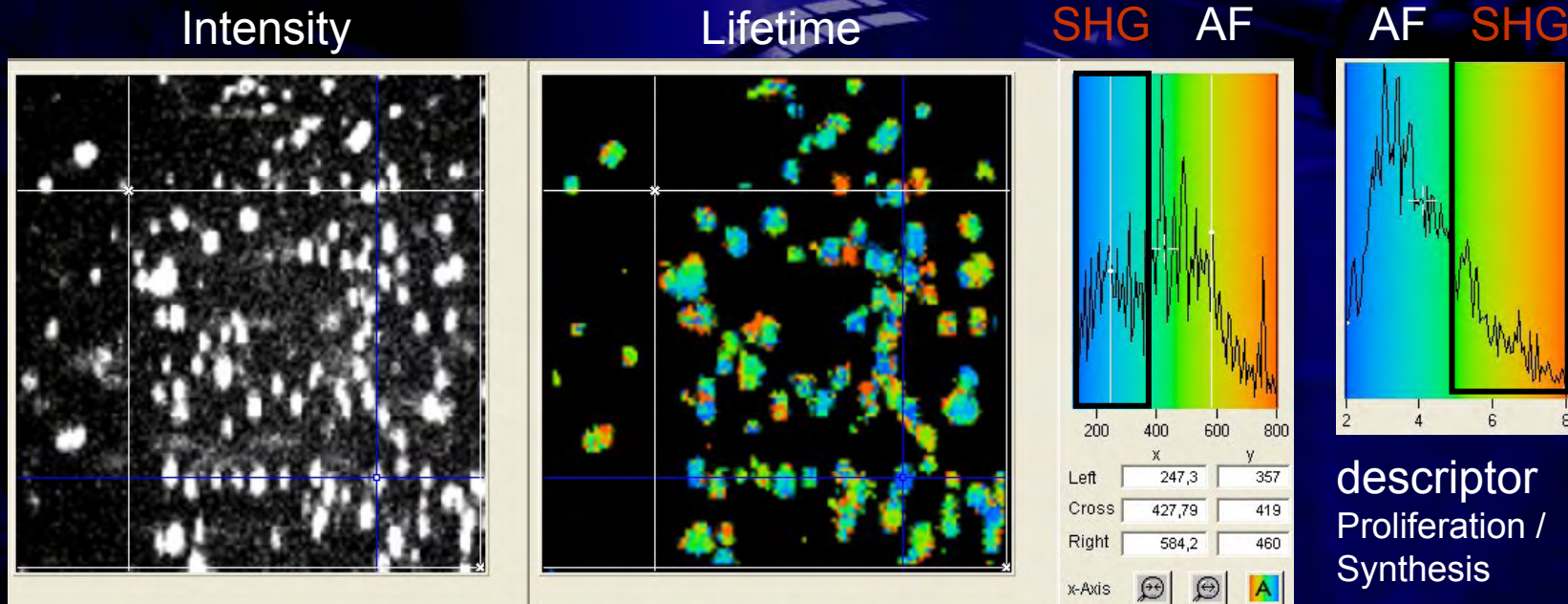
Fluorescence Decay Curves



*Mean Fluorescence lifetime (τ_m)
 Color coded image*

• SHG - TCSPC imaging : **distribution of first component**

MSC in sponge
TGF-BMP



descriptor
Proliferation /
Synthesis

t_1
SHG collagen
diffused

t_2
AF cells
reflected

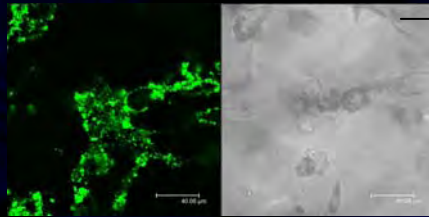
Multiexponential Decay

Components: 3

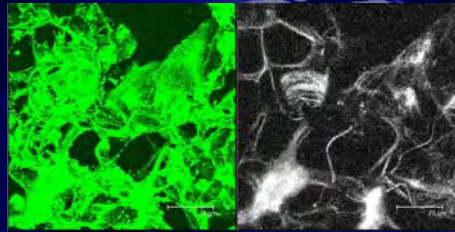
a1[%]	59.8	<input type="checkbox"/>
t1[ps]	320	<input type="checkbox"/> Fix
a2[%]	31.1	<input type="checkbox"/>
t2[ps]	1952.5	<input type="checkbox"/> Fix
a3[%]	9	<input type="checkbox"/>
t3[ps]	2241.6	<input type="checkbox"/> Fix
Shift	1.00	<input type="checkbox"/> Fix
Scatter	0.002	<input type="checkbox"/> Fix
Offset	4	<input type="checkbox"/> Fix

• SHG Multimodalities Imaging : **Limitations in Microscopy ?**

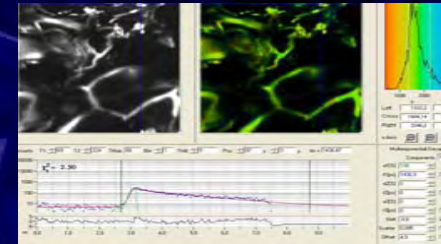
MSC in Col1 sponge – Mab anti-Col2 (alexa-488)
Coll. A.Pinzano – C. Henrionnet – P. Gillet



Fluo 2D
LT



Fluo3D
SHG



Index
SHG/AF

Limitations for medical applications : labelling / cutting / depth penetration

- Destructive assay for deep imaging or (cutting)
- Invasive protocol for labelling : exogenous probes, natural protein in tissue (elastine, collagene)
- New advances : Functional Imaging (physiological condition)



Multiscaled imaging : Macroscopy

• New advanced Macroscopy : **Confocal - Multiphoton-SHG**



CLSM

+



Macroscopy

=

Confocal Macroscopy



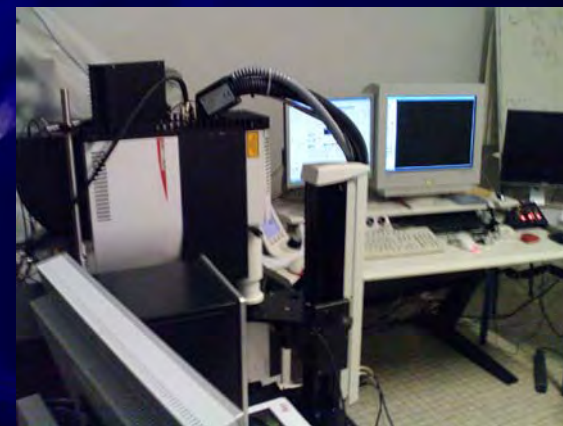
Multiphoton Laser

+



**Confocal
Macroscopy**

=



Macroscopy Multiphoton

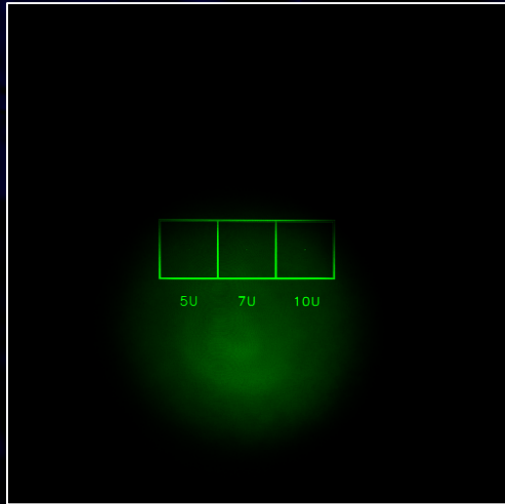


FLIM

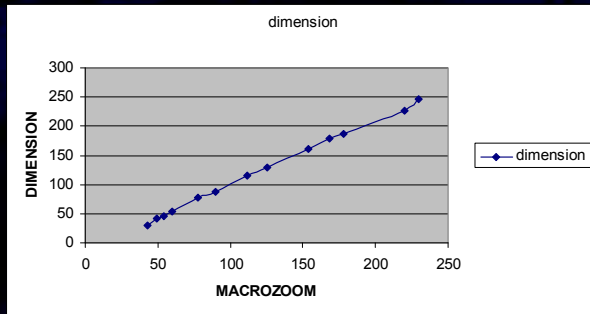
SHG

- Metrology : **MacroConfocal-MP**
- Appel à projet CNRS 2008-2009 - prototype PTIBC

488 nm

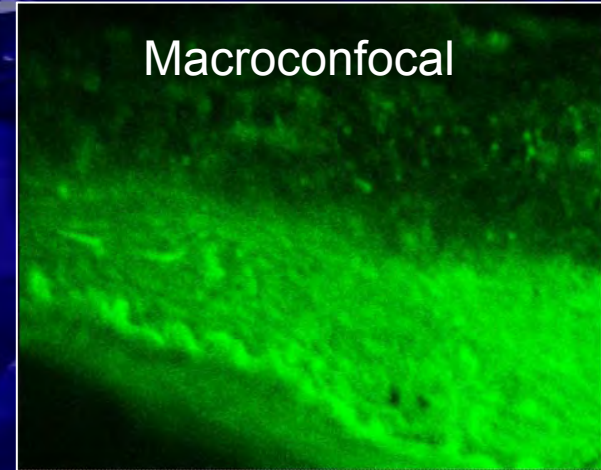


Macrozoom linearity – (20µm-5cm)



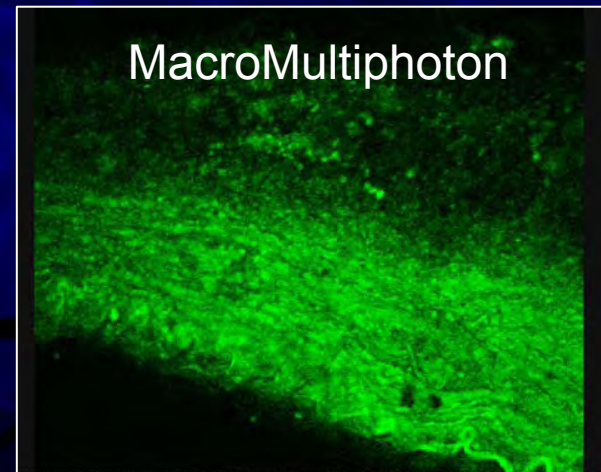
Profil Macro 2x
BE0, ZC 32, ZM 575

Rabbit carotid (autofluorescence)



Macroconfocal

488nm

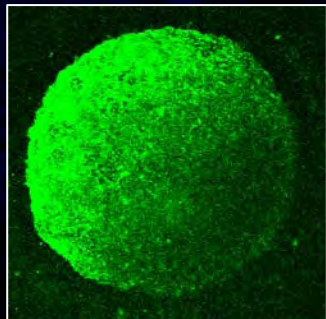


MacroMultiphoton

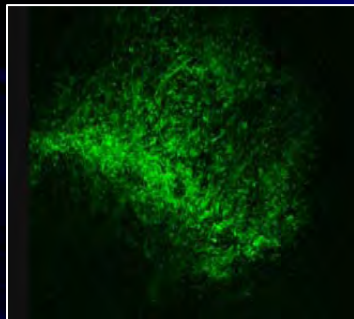
800 nm

- MacroTCSPC-SHG : Col synthesis by MSC in sponge

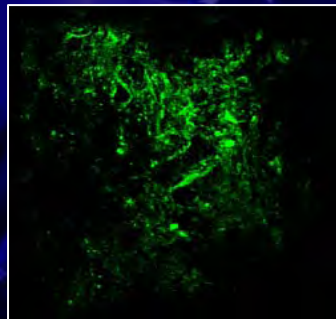
Free Cell (pure col1)



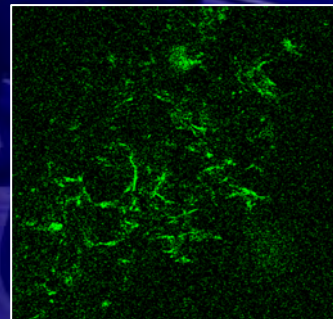
7 mm
Zoom 1,8
Macro 240x



500 μm
Zoom 18x
Macro 340x

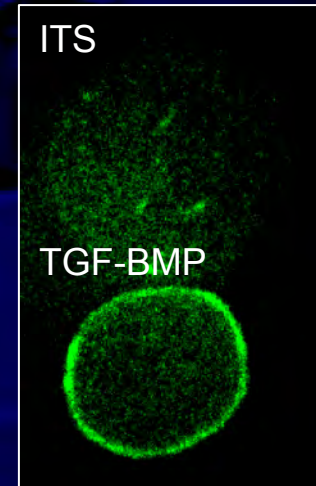


46 μm
Zoom 32x
Macro 545x



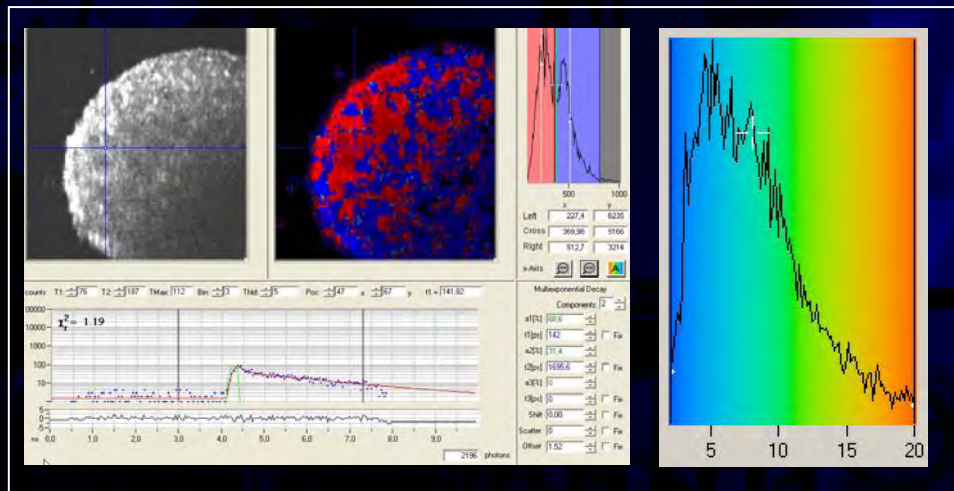
46 μm
Zoom 32x
Macro 545x

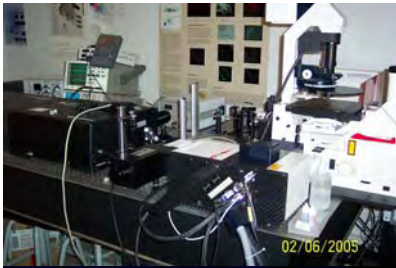
MSC



Zoom Macro 27x

Sponge of Col1





PTIBC-IBISA

UMR 7561 CNRS-Université Henri Poincaré Nancy I
Faculté de Médecine, B.P. 184
F 54505 Vandoeuvre-lès-Nancy cedex – France
D.Dumas, S. Hupont, JF. Stoltz, P. Gillet
Directeur : Jacques Magdalou
FR3209 CNRS
Directeur : Patrick Netter

Aknowledgement

C. Henrionnet
A. Pinzano
S. Hupont

CG 54
Région Lorraine
ARC
IBISA
FRM

GDR2588CNRS
RTmfm CNRS

CNRS (MRCT)
- IGBMC (Strasbourg)
- Ecole Polytechnique Palaiseau

